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(54) Title: CLONING AND RIFAMILY	ECOMBINANT PRO	ODUC	TION OI	RECEPTOR(S) OF THE	ACTIVIN/TGF-β SUPER-
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(57) Abstract					

In accordance with the present invention, there are provided novel receptor proteins characterized by having the following domains, reading from the N-terminal end of said protein: an extracellular, ligand-binding domain, a hydrophobic, trans-membrane domain, and an intracellular, receptor domain having serine kinase-like activity. The invention receptors optionally further comprise a second hydrophobic domain at the amino terminus thereof. The invention receptor proteins are further characterized by having sufficient binding affinity for at least one member of the activin/TGF- β superfamily of polypeptide growth factors such that concentrations of \leq 10 nM of said polypeptide growth factor occupy \geq 50 % of the binding sites of said receptor protein. A presently preferred member of the invention superfamily of receptors binds specifically to activins, in preference to inhibins, transforming growth factor- β , and other non-activin-like proteins. DNA sequences encoding such receptors, assays employing same, as well as antibodies derived therefrom, are also disclosed.

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CLONING AND RECOMBINANT PRODUCTION OF RECEPTOR(S) OF THE ACTIVIN/TGF-β SUPERFAMILY

ACKNOWLEDGEMENT

This invention was made with Government support under Grant Numbers HD 13527 and DK 26741, awarded by the National Institutes of Health. The Government has certain rights in this invention.

FIELD OF THE INVENTION

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The present invention relates to receptor proteins, DNA sequences encoding same, and various uses therefor.

15 BACKGROUND OF THE INVENTION

Activins are dimeric proteins which have the ability to stimulate the production of follicle stimulating hormone (FSH) by the pituitary gland. Activins share a common subunit with inhibins, which inhibit FSH secretion.

Activins are members of a superfamily of polypeptide growth factors which includes the inhibins, the transforming growth factors-B (TGF-B), Mullerian duct inhibiting substance, the Drosophila decapentaplegic peptide, several bone morphogenetic proteins, and the Vg-related peptides.

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As a result of their extensive anatomical distribution and multiple biological actions, members of this superfamily of polypeptide growth factors are believed to be involved in the regulation of numerous biological processes. Activin, for example, is involved in the proliferation of many tumor cell lines, the control of secretion and expression of the anterior pituitary hormones (e.g., FSH, GH and ACTH), neuron survival, hypothalamic oxytocin secretion, erythropoiesis, placental and gonadal steroidogenesis, early embryonic development, and the like.

Other members of the activin/TGF-B superfamily of polypeptide growth factors are involved in the regulation of cell function and cell proliferation for numerous cell types, in adults and embryos. For example, cells which are subject to regulation by one or more members of the activin/TGF-B superfamily of polypeptide growth factors include mesenchymal cells, muscle cells, skeletal cells, immune cells, hematopoietic cells, steroidogenic cells, endothelial cells, liver cells, epithelial cells, and the like.

Chemical cross-linking studies with a number of cell types suggests that multiple binding sites (i.e., receptors) exist on the surface of cells. However, little is known about the structure of these receptors, or about the second messenger signalling systems that they employ. It would be desirable, therefore, if the nature of these poorly characterized receptor proteins could be more fully understood.

BRIEF DESCRIPTION OF THE INVENTION

In accordance with the present invention, we have identified and characterized members of a new superfamily of receptor proteins which comprise three distinct domains: an extracellular, ligand-binding domain, a hydrophobic,

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trans-membrane domain, and an intracellular, receptor domain having serine kinase-like activity.

Also provided are DNAs encoding the above-5 described receptor proteins, and antibodies thereto, as well as bioassays, therapeutic compositions containing such proteins and/or antibodies, and applications thereof.

The DNAs of the invention are useful as probes for the identification of additional members of the invention superfamily of receptor proteins, and as coding sequences which can be used for the recombinant expression of the invention receptor proteins, or functional fragments thereof. The invention receptor proteins, and antibodies thereto, are useful for the diagnosis and therapeutic management of carcinogenesis, wound healing, disorders of the immune, reproductive, or central nervous systems, and the like.

20 BRIEF DESCRIPTION OF THE FIGURES

Figure 1 is a schematic diagram of receptors of the invention and the various domains thereof.

Figure 2 outlines the strategy used for expression cloning of a receptor of the activin/TGF- β receptor superfamily.

Figure 3 is a schematic of two mouse activin receptor clones. The top line of the figure is a restriction map, in kb, of mActR1 and mActR2, with numbering starting from bp 1 of mActR2. The dotted line in the figure represents 5' untranslated sequences present only in mActR1. The middle lines present a schematic representation of two activin receptor cDNA clones. Boxes represent coding sequences——black is the signal peptide, white is the extracellular ligand-binding domain, gray is

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the transmembrane, and the intracellular kinase domain is hatched. Amino acids are numbered beneath the schematics.

Figure 4 presents a comparison between activin receptor and daf-1 [a C. elegans gene encoding a putative receptor protein kinase (with unknown ligand); see Georgi, et al., Cell 61: 635-645 (1990)]. Conserved residues between the activin receptor and daf-1 are highlighted; conserved kinase domain residues are designated with an "*".

Figure 5A summarizes results of ¹²⁵I activin A binding to COS cells transfected with pmActR1. Binding was competed with unlabeled activin A. For the runs reported 15 herein, total binding was 4.6% of input cpm, non-specific binding was 0.9% of input cpm, and therefore the specific binding was 3.7% of input cpm. Data are shown as % specific binding, normalized to 100%. The inset presents a Scatchard analysis of the data [Ann. NY Acad. Sci. 51: 20 660-672 (1979)].

Figure 5B summarizes results of 125I activin A binding to COS cells transfected with pmActR2. Binding was competed with unlabeled factors as indicated in the figure.

25 For the runs reported herein, total binding was 3.4% of input cpm, non-specific binding was 0.9% of input cpm, and therefore the specific binding was 2.5% of input cpm. Data are shown as % specific binding, normalized to 100%.

Figure 6 is a phylogenetic tree, comparing the relationship of the activin receptor kinase domain to other protein kinases. To construct the tree, the catalytic domains of representative sequences were empirically aligned and evolutionary relatedness was calculated using an algorithm designed by Fitch and Margoliash [Science 155: 279-284 (1967)], as implemented by Feng and Doolittle [J. Mol. Evol. 25: 351-360 (1987)]. Known subfamilies of

kinases are indicated in the figure. For those sequences that had similarity scores (i.e., a relative sequence identity) of at least 4 standard deviations above the mean (in comparison with all other known kinase sequences), the percent identity with the activin receptor is indicated. For further detail on kinase sequences, the reader is referred to Hanks and Quinn, Meth. Enzymol. 200: 38-62 (1991).

10 <u>DETAILED DESCRIPTION OF THE INVENTION</u>

In accordance with the present invention, there is provided a novel superfamily of receptor protein(s) characterized by having the following domains, reading from the N-terminal end of said protein:

an extracellular, ligand-binding domain,
a hydrophobic, trans-membrane domain, and
an intracellular domain having serine kinase-like
activity.

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The novel receptor protein(s) of the invention optionally further comprise a second hydrophobic domain at the amino terminus thereof.

As employed herein, the phrase "extracellular, ligand-binding domain" refers to that portion of receptors of the invention which has a high affinity for ligand, and which, when associated with a cell, resides primarily outside of the cell membrane. Because of its location, this domain is not exposed to the processing machinery present within the cell, but is exposed to all components of the extracellular medium. See Figure 1.

As employed herein, the phrase "hydrophobic, 35 trans-membrane domain" refers to that portion of receptors of the invention which traverses the cell membrane, and serves as a "bridge" between the extracellular and

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intracellular domains of the receptor. The hydrophobic nature of this domain serves to anchor the receptor to the cell membrane. See Figure 1.

As employed herein, the phrase "intracellular domain having serine kinase-like activity" refers to that portion of receptors of the invention which resides within the cytoplasm, and which embodies the catalytic functionality characteristic of all receptors of the invention. See Fig 1.

The optional second hydrophobic domain, positioned at the amino terminus of receptors of the invention, comprises a secretion signal sequence which promotes the intracellular transport of the initially expressed receptor protein across the Golgi membrane. See Figure 1.

Members of the invention superfamily of receptors

20 can be further characterized as having sufficient binding
affinity for at least one member of the activin/TGF-ß
superfamily of polypeptide growth factors such that
concentrations of ≤ 10 nM of said polypeptide growth factor
occupy ≥ 50% of the binding sites of said receptor protein.

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Binding affinity (which can be expressed in terms of association constants, Ka, or dissociation constants, Kd) refers to the strength of interaction between ligand and receptor, and can be expressed in terms of the concentration of ligand necessary to occupy one-half (50%) of the binding sites of the receptor. A receptor having a high binding affinity for a given ligand will require the presence of very little ligand to become at least 50% bound (hence the Kd value will be a small number); conversely, receptor having a low binding affinity for a given ligand will require the presence of high levels of ligand to become 50% bound (hence the Kd value will be a large

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number).

Reference to receptor protein "having sufficient binding affinity such that concentrations of 5 polypeptide growth factor less than or equal to 10 nm (i.e., ≤ 10 nM) occupy ≥ 50% (i.e., greater than or equal to one-half) of the binding sites of said receptor protein" means that ligand (i.e., polypeptide growth factor) concentration(s) of no greater than about 10 nM are 10 required in order for the ligand to occupy at least 50% of the active sites of said receptor, with much lower ligand concentrations typically being required. preferred receptors of the present invention have a binding affinity such that ligand concentration(s) in the range of 15 only about 100 - 500 pM are required in order to occupy (or bind to) at least 50% of the receptor binding sites.

Members of the invention superfamily of receptors can be divided into various subclasses, based on the approximate size of the crosslinked complexes obtained when radiolabeled activin is chemically crosslinked to cell extracts [see, for example, Example VI below, or Mathews and Vale in Cell 65:973-982 (1991)]. Type I activin/TGF-β receptors are those which form a crosslinked complex of about 65 kD with activin; Type II receptors are those which form a crosslinked complex of about 80-85 kD with activin; while Type III, Type IV and the like receptors are those which form crosslinked complexes with activin having molecular weights greater than about 100 kD.

30

Each member of a given subclass is related to other members of the same subclass by the high degree of homology (e.g., >80% overall amino acid homology; frequently having >90% overall amino acid homology) between such receptors; whereas members of a given subclass differ from members of a different subclass by the lower degree of homology (e.g., at least about 30% up to 80% overall amino

acid homology; with in the range of about 40% up to 90% amino acid homology specifically in the kinase domains thereof) between such receptors. Typically, related receptors have at least 50% overall amino acid homology; with at least about 60% amino acid homology in the kinase domains thereof. Preferably, related receptors are defined as those which have at least 60% overall amino acid homology; with at least about 70% amino acid homology in the kinase domains thereof.

10

Based on the above criteria, the receptors described herein are designated Type II receptors, with the first discovered Type II receptor (i.e., the mouse-derived activin receptor) being designated ActRII, while subsequently identified Type II receptors which are not homologs of ActRII (because while clearly related by size and some sequence homology, they differ sufficiently to be considered as variants of ActRII), are designated ActRIIB, ActRIIC, etc.

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Presently preferred members of the invention superfamily of receptors are further characterized by having a greater binding affinity for activins than for inhibins. Such receptors are frequently also observed to 25 have:

substantially no binding affinity for transforming growth factors-B, and

substantially no binding affinity for non-activin-like proteins or compounds.

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Additional members of the invention superfamily of receptors are further characterized by having a greater binding affinity for inhibins than for activins or TGF-Bs.

Additional members of the invention superfamily of receptors are further characterized by having a greater binding affinity for TGF-Bs than for activins or inhibins.

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As employed herein, "activin" refers to activin A (a homodimer of two inhibin β_A subunits), activin B (a homodimer of two inhibin β_B subunits), activin AB (a heterodimer composed of one inhibin β_A subunit and one inhibin β_B subunit); "inhibin" refers to inhibin A (composed of the inhibin α subunit and an inhibin β_A subunit), inhibin B (composed of the inhibin α subunit and an inhibin β_B subunit); "transforming growth factor B or TGF-B" refers to TGF-B1 (a homodimer of two TGF-B1 subunits), TGF-B2 (a homodimer of two TGF-B3 subunits), TGF-B4 (a homodimer of two TGF-B4 subunits), TGF-B5 (a homodimer of two TGF-B5 subunits), TGF-B5 (a heterodimer of one TGF-B1 subunit and one TGF-B2 subunit), and the like.

15

Transforming growth factors- β (TGF- β s) are members of the activin/TGF- β superfamily of polypeptide growth factors. TGF- β s are structurally related to activins, sharing at least 20-30% amino acid sequence homology therewith. TGF- β s and activins have a substantially similar distribution pattern of cysteine residues (or substitution) throughout the peptide chain. Furthermore, both polypeptides, in their active forms, are dimeric species.

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As employed herein, the term "non-activin-like" proteins refers to any protein having essentially no structural similarity with activins (as defined broadly herein).

30

Preferred members of the invention superfamily of receptors comprise those having in the range of about 500 amino acids, and are further characterized by having the following designated sizes for each of the domains thereof, reading from the N-terminal end of said receptor:

the extracellular, ligand-binding domain preferably will have in the range of about 114-118

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amino acids,

the hydrophobic, trans-membrane domain preferably will have in the range of about 23-28 amino acids, beginning at the carboxy terminus of the extracellular domain, and

the intracellular domain having kinase-like activity preferably will have in the range of about 345-360 amino acids, beginning at the carboxy terminus of the hydrophobic, trans-membrane domain.

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Receptors of the invention optionally further comprise a second hydrophobic domain having in the range of about 16-30 amino acids at the extreme amino terminus thereof (i.e., at the amino terminus of the extracellular, ligand-binding domain). This domain is a secretion signal sequence, which aids the transport of invention receptor(s) across the cell membrane. Exemplary secretion signal sequences include amino acids 1-19 of Sequence ID No. 1, amino acids 1-20 of Sequence ID No. 3, and the like. Such secretion signal sequences can be encoded by such nucleic acid sequences as nucleotides 71-127 of Sequence ID No. 1, nucleotides 468-527 of Sequence ID No. 3, and the like.

Members of the invention superfamily of receptors

25 can be obtained from a variety of sources, such as, for
example, pituitary cells, placental cells, hematopoietic
cells, brain cells, gonadal cells, liver cells, bone cells,
muscle cells, endothelial cells, epithelial cells,
mesenchymal cells, kidney cells, and the like. Such cells

30 can be derived from a variety of organisms, such as, for
example, human, mouse, rat, ovine, bovine, porcine, frog,
chicken, fish, mink, and the like.

Presently preferred amino acid sequences encoding 35 receptor proteins of the invention include the sequence set forth in Sequence ID No. 2 (which represents a mouse activin receptor amino acid sequence), a modified form of

Sequence ID No. 2 wherein the arginine at residue number 39 is replaced by a lysine, the isoleucine at residue number 92 is replaced by a valine, and the glutamic acid at residue number 288 is replaced by a glutamine (which 5 modified form of Sequence ID No. 1 is referred to hereinafter as "Sequence ID No. 1'", and represents a human activin receptor amino acid sequence), and the sequence set forth as Sequence ID No. 4 (which represents a Xenopus activin receptor amino acid sequence), as well 10 functional, modified forms thereof. Those of skill in the art recognize that numerous residues of the above-described sequences can be substituted with other, chemically, sterically and/or electronically similar residues without substantially altering the biological activity of the 15 resulting receptor species.

In accordance with another embodiment of the present invention, there is provided a soluble, extracellular, ligand-binding protein, further 20 characterized by:

having sufficient binding affinity for at least one member of the activin/TGF-B superfamily of polypeptide growth factors such that concentrations of \leq 10 nM of said polypeptide growth factor occupy \geq 50% of the binding sites on said receptor protein, and

having at least about 30% sequence identity with respect to:

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the sequence of amino acids 20-134 set forth in Sequence ID No. 2;

the sequence of amino acids 20-134 set forth in Sequence ID No. 2, wherein the arginine residue at position number 39 is replaced by a lysine, and the isoleucine at residue number 92 is replaced by a valine; or

the sequence of amino acids 21-132 set forth in Sequence ID No. 4.

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Presently preferred soluble, extracellular, ligand-binding proteins contemplated by the present invention can be further characterized by having at least about 50% sequence identity with respect to:

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the sequence of amino acids 20-134 set forth in Sequence ID No. 2;

the sequence of amino acids 20-134 set forth in Sequence ID No. 2, wherein the arginine residue at position number 39 is replaced by a lysine, and the isoleucine at residue number 92 is replaced by a valine; or

the sequence of amino acids 21-132 set forth in Sequence ID No. 4;

with the presently most preferred soluble, extracellular,
ligand-binding proteins having at least about 80% sequence
identity with respect to the above-referenced fragments of
Sequence ID Nos. 2 or 4.

Members of the class of soluble, ligand-binding 20 proteins contemplated by the present invention may be divided into various subclasses, as previously described, wherein members of one subclass may have a greater binding affinity for activins than for inhibins and/or TGF-Bs; or alternatively, members of another subclass may have a 25 greater binding affinity for inhibins than for activins and/or TGF-Bs; or alternatively, members of yet another subclass may have a greater binding affinity for TGF-Bs It is, of course, than for activins and/or inhibins. understood by those of skill in the art, that members of 30 more than one subclass may have a greater binding affinity for one member of the activin/TGF-B superfamily of polypeptide growth factors, relative to other members of the superfamily.

Presently preferred soluble, extracellular, ligand-binding proteins of the present invention are further characterized by:

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having a greater binding affinity for activins than for inhibins,

having substantially no binding affinity for transforming growth factors-8, and

5 having substantially no binding affinity for non-activin-like proteins.

Presently preferred soluble, extracellular, ligand-binding proteins of the present invention typically comprise in the range of about 114-118 amino acids.

Especially preferred soluble, extracellular, ligand-binding proteins of the invention are those having substantially the same amino acid sequence as that set forth as:

residues 20-134 of Sequence ID No. 2;

residues 20-134 of Sequence ID No. 2, wherein the arginine residue at position number 39 is replaced by a lysine, and the isoleucine at residue number 92 is replaced by a valine; or

residues 21-132 of Sequence ID No. 4.

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As employed herein, the term "substantially the same amino acid sequence" refers to amino acid sequences

25 having at least about 80% identity with respect to the reference amino acid sequence, and will retain comparable functional and biological properties characteristic of the protein encoded by the reference amino acid. Preferably, proteins having "substantially the same amino acid sequence" will have at least about 90% amino acid identity with respect to the reference amino acid sequence; with greater than about 95% amino acid sequence identity being especially preferred.

The above-described soluble proteins can be employed for a variety of therapeutic uses, e.g., to block receptors of the invention from affecting processes which

the receptors would otherwise mediate. The presence of the soluble proteins of the invention will compete with functional ligand for the receptor, preventing the formation of a functional receptor-ligand complex, thereby blocking the normal regulatory action of the complex.

In accordance with yet another embodiment of the present invention, there are provided antibodies generated against the above-described soluble proteins and receptor proteins. Such antibodies can be employed for diagnostic applications, therapeutic applications, and the like. Preferably, for therapeutic applications, the antibodies employed will be monoclonal antibodies.

The above-described antibodies can be prepared employing standard techniques, as are well known to those of skill in the art, using the invention receptor proteins as antigens for antibody production.

In accordance with still another embodiment of the present invention, there are provided methods for modulating the transcription trans-activation of receptor(s) of the invention by contacting said receptor(s) with a modulating, effective amount of the above-described antibodies.

The soluble proteins of the invention, and the antibodies of the invention, can be administered to a subject employing standard methods, such as, for example, by intraperitoneal, intramuscular, intravenous, or subcutaneous injection, implant or transdermal modes of administration, and the like. In addition, methods such as transfection with viral or retroviral vectors encoding the invention compositions. One of skill in the art can readily determine dose forms, treatment regiments, etc, depending on the mode of administration employed.

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In accordance with a further embodiment of the present invention, there are provided DNA sequences which encode the above-described soluble proteins and receptor proteins. Optionally, such DNA sequences, or fragments thereof, can be labeled with a readily detectable substituent (to be used, for example, as a hybridization probe).

The above-described receptor(s) can be encoded by numerous DNA sequences, e.g., a DNA sequence having a contiguous nucleotide sequence substantially the same as:

nucleotides 128 - 1609 of Sequence ID No. 1 (which encodes a mouse activin receptor);

variations of nucleotides 128 - 1609 of Sequence ID No. 1, wherein the codon for residue number 39 of the encoded amino acid codes for lysine, the codon for residue number 92 of the encoded amino acid codes for valine, and the codon for residue number 288 of the encoded amino acid encodes glutamine (which encodes a human activin receptor);

nucleotides 528 - 1997 of Sequence ID No. 3 (which encodes a Xenopus activin receptor); or

variations of any of the above sequences which encode the same amino acid sequences, but employ different codons for some of the amino acids.

As employed herein, the term "substantially the same as" refers to DNA having at least about 70% homology with respect to the nucleotide sequence of the DNA fragment with which subject DNA is being compared. Preferably, DNA "substantially the same as" a comparative DNA will be at least about 80% homologous to the comparative nucleotide sequence; with greater than about 90% homology being especially preferred.

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Another DNA which encodes a receptor of the invention is one having a contiguous nucleotide sequence

substantially the same as:

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nucleotides 71 - 1609 of Sequence ID No. 1 (which encodes a precursor-form of a mouse activin receptor);

variations of nucleotides 71 - 1609 of Sequence ID No. 1, wherein the codon for residue number 39 of the encoded amino acid codes for lysine, the codon for residue number 92 of the encoded amino acid codes for valine, and the codon for residue number 288 of the encoded amino acid encodes glutamine (which encodes a precursor-form of a human activin receptor);

nucleotides 468 - 1997 of Sequence ID No. 3 (which encodes a precursor form of a Xenopus activin receptor); or

variations of any of the above sequences which encode the same amino acid sequences, but employ different codons for some of the amino acids.

Yet another DNA which encodes the above-described receptor is one having a contiguous nucleotide sequence substantially the same as set forth in Sequence ID No. 1, Sequence ID No. 1' or Sequence ID No. 3.

In accordance with a further embodiment of the present invention, the receptor-encoding cDNAs can be 25 employed to probe library(ies) (e.g., cDNA, genomic, and the like) for additional sequences encoding novel receptors Such screening is of the activin/TGF- β superfamily. initially carried out under low-stringency conditions, which comprise a temperature of less than about 42°C, a 30 formamide concentration of less than about 50%, and a moderate to low salt concentration. Presently preferred conditions for such screening comprise a temperature of about 37°C, a formamide concentration of about 20%, and a salt concentration of about 5X standard saline citrate 35 (SSC; 20X SSC contains 3M sodium chloride, 0.3M sodium Such conditions will allow the citrate, pH 7.0). identification of sequences which have a substantial degree

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of similarity with the probe sequence, without requiring perfect homology for the identification of a stable hybrid. The phrase "substantial similarity" refers to sequences which share at least 50% homology. Preferably, hybridization conditions will be selected which allow the identification of sequences having at least 70% homology with the probe, while discriminating against sequences which have a lower degree of homology with the probe.

In accordance with yet another embodiment of the present invention, there is provided a method for the recombinant production of receptor(s) of the invention by expressing the above-described DNA sequences in suitable host cells.

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The use of a wide variety of recombinant organisms has been described for the production of peptides. One of skill in the art can readily determine suitable hosts (and expression conditions) for use in the recombinant production of the peptides of the present invention. Yeast hosts, bacterial hosts, mammalian hosts, and the like can be employed. Regulatory sequences capable of controlling the expresseion of invention peptides are well known for each of these host systems, as are growth conditions under which expression occurs.

In accordance with a further embodiment of the present invention, there is provided a binding assay employing receptors of the invention, whereby a large number of compounds can be rapidly screened to determine which compounds, if any, are capable of binding to the receptors of the invention. Then, more detailed assays can be carried out with those compounds found to bind, to further determine whether such compounds act as agonists or antagonists of invention receptors.

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Another application of the binding assay of the invention is the assay of test samples (e.g., biological fluids) for the presence or absence of members of the activin/TGF-B superfamily of polypeptide growth factors.

5 Thus, for example, serum from a patient displaying symptoms related to pathway(s) mediated by members of the activin/TGF-B superfamily of polypeptide growth factors can be assayed to determine if the observed symptoms are perhaps caused by over- or under-production of such polypeptide growth factor.

The binding assays contemplated by the present invention can be carried out in a variety of ways, as can readily be identified by one of skill in the art. For example, competitive binding assays can be employed, as well as radioimmunoassays, ELISA, ERMA, and the like.

In accordance with a still further embodiment of the present invention, there are provided bioassays for 20 evaluating whether test compounds are capable of acting as agonists or antagonists of receptor(s) of the present invention.

The bioassays of the present invention involve evaluating whether test compounds are capable of acting as either agonists or antagonists for members of the invention superfamily of receptors, or functional modified forms of said receptor protein(s). The bioassay for evaluating whether test compounds are capable of acting as agonists comprises:

(a) culturing cells containing:

DNA which expresses said receptor protein(s) or functional modified forms of said receptor protein(s), and

DNA encoding a hormone response element operatively linked to a reporter gene; wherein said culturing is carried out in the

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presence of at least one compound whose ability to induce transcription activation activity of receptor protein is sought to be determined, and thereafter

5 (b) monitoring said cells for expression of the product of said reporter gene.

The bioassay for evaluating whether test compounds are capable of acting as antagonists for 10 receptor(s) of the invention, or functional modified forms of said receptor(s), comprises:

(a) culturing cells containing:

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DNA which expresses said receptor protein(s), or functional modified forms of said receptor protein(s), and

DNA encoding a hormone response element operatively linked to a reporter gene wherein said culturing is carried out in the presence of:

increasing concentrations of at least one compound whose ability to inhibit transcription activation of said receptor protein(s) is sought to be determined, and

a fixed concentration of at least one agonist for said receptor protein(s), or functional modified forms of said receptor protein(s); and thereafter

(b) monitoring in said cells the level of expression of the product of said reporter gene as a function of the concentration of said compound, thereby indicating the ability of said compound to inhibit activation of transcription.

Host cells contemplated for use in the 35 bioassay(s) of the present invention, include CV-1 cells, COS cells, and the like; reporter and expression plasmids employed typically also contain the origin of replication

of SV-40; and the reporter and expression plasmids employed also typically contain a selectable marker.

The hormone response element employed in the bioassay(s) of the present invention can be selected from, for example, mouse mammary tumor virus long terminal repeat (MTV LTR), mammalian growth hormone promoter, and the reporter gene can be selected from chloramphenicol acetytransferase (CAT), luciferase, B-galactosidase, and the like.

The cells can be monitored for the level of expression of the reporter gene in a variety of ways, such as, for example, by photometric means [e.g., by colorimetry (with a colored reporter product such as B-galactosidase), by fluorescence (with a reporter product such as luciferase), etc], by enzyme activity, and the like.

Compounds contemplated for screening in accordance with the invention bioassays include activin- or $TGF-\beta$ -like compounds, as well as compounds which bear no particular structural or biological relatedness to activin or $TGF-\beta$.

As employed herein, the phrase "activin- or $TGF-\beta$ -like compounds" includes substances which have a substantial degree of homology (at least 20% homology) with the amino acid sequences of naturally occurring mammalian inhibin alpha and β_A or β_B chains (either singly or in any combination) as well as alleles, fragments, homologs or derivatives thereof which have substantially the same qualitative biological activity as mammalian inhibin, activin, or $TGF-\beta$. Examples of activin- or $TGF-\beta$ -like compounds include activin A (a homodimer of two inhibin β_A subunits), activin B (a homodimer of two inhibin β_B subunits), activin AB (a heterodimer composed of one inhibin β_A subunit and one inhibin β_B subunit), inhibin A

(composed of the inhibin α subunit and an inhibin B_A subunit), inhibin B (composed of the inhibin α subunit and an inhibin B_B subunit), TGF-B1 (a homodimer of two TGF-B1 subunits), TGF-B2 (a homodimer of two TGF-B2 subunits), TGF-B3 (a homodimer of two TGF-B3 subunits), TGF-B4 (a homodimer of two TGF-B4 subunits), TGF-B5 (a homodimer of two TGF-B5 subunits), TGF-B1.2 (a heterodimer of one TGF-B1 subunit and one TGF-B2 subunit), and the like.

Examples of compounds which bear no particular structural or biological relatedness to activin or TGF-β, but which are contemplated for screening in accordance with the bioassays of the present invention, include any compound that is capable of either blocking the action of the invention receptor peptides, or promoting the action of the invention receptor peptides, such as, for example, alkaloids and other heterocyclic organic compounds, and the like.

The method employed for cloning the receptor(s) of the present invention involves expressing, in mammalian cells, a cDNA library of any cell type thought to respond to members of the activin/TGF-B superfamily of polypeptide growth factors (e.g., pituitary cells, placental cells, fibroblast cells, and the like). Then, the ability of the resulting mammalian cells to bind a labeled receptor ligand (i.e., a labeled member of the activin/TGF-B superfamily of polypeptide growth factors) is determined. Finally, the desired cDNA insert(s) are recovered, based on the ability of that cDNA, when expressed in mammalian cells, to induce (or enhance) the binding of labeled receptor ligand to said cell.

In addition to the above-described applications
of the receptor proteins and DNA sequences of the present
invention, the receptor or receptor-encoding compositions
of the invention can be used in a variety of ways. For

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example, since activin is involved in many biological processes, the activin receptor (or antibodies thereto) can be applied to the modulation of such biological processes. For example, the stimulation of FSH release by activin can either be enhanced (for example, by supplying the subject with increased amounts of the activin receptor, relative to the amount of endogenous receptor, e.g., by transfecting the subject with a tissue specific activin-encoding construct), or depressed (e.g., by administration to a subject of antibodies to the activin receptor, thereby preventing formation of activin-receptor complex, which would then act to stimulate the release of FSH). Thus, the compositions of the present invention can be applied to the control of fertility in humans, domesticated animals, and animals of commercial interest.

As another example, the effect of activin on mitosis of red and white blood cells can be modulated, for example, by administering to a subject (employing suitable means of administration) a modulating, effective amount of activin receptor (which would enhance the ability of activin present in the cell to modulate mitosis). Alternatively, one could administer to a subject an antibody to the activin receptor (or a portion thereof), which would reduce the effect of activin by blocking the normal interaction between activin and activin receptor.

As additional examples of the wide utility of the invention compositions, receptors and/or antibodies of the invention can be used in such areas as the diagnosis and/or treatment of activin-dependent tumors, enhancing the survival of brain neurons, inducing abortion in livestock and other domesticated animals, inducing twinning in livestock and other domesticated animals, and so on.

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As still further examples of the wide utility of the invention compositions, agonists identified for TGF-B

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specific receptors can be used to stimulate wound healing, to suppress the growth of TGF-B-sensitive tumors, to suppress immune response (and thereby prevent rejection of transplanted organs), and the like. Antagonists or the soluble, ligand-binding domain derived from TGF-B receptors can be used to block endogenous TGF-B, thereby promoting liver regeneration and stimulating some immune responses.

It can be readily seen, therefore, that the invention compositions have utility in a wide variety of diagnostic, clinical, veterinary and research applications.

The invention will now be described in greater detail by reference to the following non-limiting examples.

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EXAMPLES

Recombinant human (rh) activin A, rh activin B, and rh inhibin A were generously provided by Genentech,

20 Inc. Porcine TGF-B1 was obtained from R+D Systems.

Double-stranded DNA was sequenced by the dideoxy chain termination method using the Sequenase reagents from US Biochemicals. Comparison of DNA sequences to databases was performed using the FASTA program [Pearson and Lipman, Proc. Natl. Acad. Sci. USA 85: 2444-2448 (1988)].

EXAMPLE I

Construction and Subdivision of cDNA Library

30

Polyadenylated RNA was prepared from AtT20 cells using the Fast Track reagents from InVitrogen. cDNA was commercially synthesized and ligated into the plasmid vector pcDNA1 using non-palindromic BstXI linkers, yielding a library of approximately 5×10^6 primary recombinants. The unamplified cDNA library was plated at 1000 clones per 100 mm plate, then scraped off the plates, frozen in glycerol

and stored at -70°.

Activin suppresses adrenocorticotrophic hormone (ACTH) secretion by both primary anterior pituitary cell 5 cultures [Vale et al., Nature 321: 776-779 (1986)] and Because AtT20 cells AtT20 mouse corticotropic cells. possess activin receptors indistinguishable from those on other cell types (based on binding affinity measurements with activin A), these cells were chosen to be the source 10 of cDNA for transfection. A cDNA library of approximately 5x106 independent clones from AtT20 cells was constructed in the mammalian expression vector, pcDNA1, and screened using an expression cloning approach [Gearing et al., EMBO J. 8, 3667-3676 (1989)] based on the ability to detect activin 15 binding to single transfected cells. The library was divided into pools of 1000 clones, DNA was prepared from each pool of clones and transiently transfected into COS cells, and the cells screened for the capacity to bind iodinated activin A. Binding was assessed by performing 20 the transfections and binding reactions directly on chambered microscope slides, then dipping the slides in emulsion and analyzing them under photographic Cells which had been transfected with an microscope. activin receptor cDNA, and consequently bound radioactive 25 activin, were covered with silver grains. DNA from pools of clones were analyzed either singly or in groups of three. Of 300 pools (approximately 300,000 clones) assayed in this manner, one group of three generated two positive cells when transfected into COS cells. The positive pool 30 (#64) was identified by transfecting and analyzing DNA from each pool of 1000 singly, and then was further fractionated until a single clone (pmActR1) was purified which generated >104 positive cells after transfection (see Table 1).

Table 1
Purification of the activin receptor clone from the AtT20 library

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5	Pool	Clones/pool	Positive cells/slide
	62,63,64	3x1000	2
	64	1000	1-3
	64-51	400	4-10
	64-51-R10;64-51-C13	20	25-40
10	pmActR1	1	>104

The total number of transfected cells capable of binding 125 I activin A in a field of 2x10⁵ COS cells was counted for pools of clones at each stage of the purification process.

pmActR1 contained a 1.7 kb insert, coding for a protein of 342 amino acids (Figure 3); however, it was incomplete on the 3' end, thus the last 17 amino acids were encoded by vector sequences. In order to obtain the entire sequence, the AtT20 library was rescreened by hybridization with the 1.6 kb SacI-PstI fragment (Figure 3). Screening 6x10⁵ colonies yielded one additional positive clone (pmActR2) which had a 2.6 kb insert and contained the entire coding sequence for the mouse activin receptor (Figure 3). The nucleic acid sequence and the deduced amino acid sequence of the insert in pmActR2 are set forth in Sequence ID No. 1.

30 <u>EXAMPLE II</u> <u>COS Cell Transfection</u>

Aliquots of the frozen pools of clones were grown overnight in 3 ml cultures of terrific broth, and mini-prep DNA prepared from 1.5 ml using the alkaline lysis method [Maniatis et al. Molecular Cloning (Cold Spring Harbor Laboratory (1982)]. 1/10 of the DNA from a mini-prep (10

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Ml of 100 Ml) was used for each transfection.

cos cells were plated on chambered 2x10⁵ microscope slides (1 chamber - Nunc) that had been coated 5 with 20 μ g/ml poly-D-lysine and allowed to attach for at Cells were subjected to DEAE-Dextran least 3 hours. mediated transfection as follows. 1.5 ml of serum-free Dulbecco's Modified Eagle's medium (DME) containing 100 mM chloroquine was added to the cells. DNA was precipitated 10 in 200 ml DME/chloroquine containing 500 mg/ml DEAE-Dextran, then added to the cells. The cells were incubated at 37° for 4 hours, then the media was removed and the cells were treated with 10% DMSO in HEPES buffered saline for 2 minutes. Fresh media was added and the cells assayed 15 3 days later. For transfections with the purified clone, $2.5 \times 10^{\circ}$ cells were transfected in 100 mm dishes with 5 μg purified DNA. The total transfection volume was 10 ml, and the DNA was precipitated in 400 μ l.

20 <u>EXAMPLE III</u> Binding Assay

Cells were washed 2x with HEPES buffered saline (HDB) containing 0.1% BSA, then incubated for 90 minutes at 25 22° in 0.5 ml HDB, 0.1% BSA containing 7x10⁵ cpm 125 activin A (approximately 7 ng, 500 pM). The cells were then washed 3X with cold HDB, fixed for 15 minutes at 22° in 2.5% glutaraldehyde/HDB and washed 2X with HDB. The chambers were then peeled off the slides, and the slides dehydrated 30 in 95% ethanol, dried under vacuum, dipped in NTB2 photographic emulsion (Kodak) and exposed in the dark at 4° Following development of the emulsion, the for 3 days. slides were dehydrated in 95% ethanol, stained with eosin and coverslipped with DPX mountiant (Electron Microscopy The slides were analyzed under darkfield 35 Sciences). illumination using a Leitz microscope.

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EXAMPLE IV Subdivision of Positive Pool

Of 300 pools screened (each pool containing about 5 1000 cDNAs), one positive pool (#64), which produced two positive cells, was identified. Bacteria from the frozen stock of this positive pool (#64) were replated at approximately 400 clones per plate, replica plates were made, and DNA was prepared from each subpool and analyzed 10 employing the binding assay described above. positive subpools were found, which generated from 4-10 positive cells per slide. The bacteria from the replica plate of one positive subpool were picked onto a grid, and DNA prepared from pools of clones representing all the rows 15 and all the columns, as described by Wong [Science 228:810-The identification of one positive row and 815 (1985)]. one positive column unambiguously identified a single clone, which when transfected yielded >104 positive cells/2x10⁵ cells.

20

EXAMPLE V Radioreceptor Assay

pmActR2, or 10⁶ untransfected COS cells, were plated in 6 well dishes and allowed to grow overnight. The cells were washed 2X with HDB, 0.1% BSA, and incubated at 22° for 90 minutes in 0.5 ml HDB, 0.1% BSA containing 100,000 cpm (approximately 1 ng, 75 pM) ¹²⁵I activin A (5 μg activin A was iodinated by chloramine T oxidation to a specific activity of 50-90 μCi/μg; iodinated activin A was purified on a 0.7x20 cm G-25 column) and varying amounts of unlabeled competitor hormone. Following binding, the cells were washed 3X with cold HDB, solubilized in 0.5 ml 0.5 N NaOH, removed from the dish and radioactivity was measured in a gamma counter. Data presented in Figure 5 are expressed as % specific binding, where 100% specific

binding is the difference between binding in the absence of competitor and binding in the presence of a 100 fold molar excess of unlabeled activin A. Binding parameters were determined using the program LIGAND [Munson P.J. and Rodbard, D., Anal. Biochem. 107:220-259 (1980)].

EXAMPLE VI Chemical Cross-linking

2x10⁶ COS cells, or 5x10⁶ AtT20 cells, were washed 10 2x with HDB, scraped off the dish, incubated for 90 minutes at 22° under constant rotation in 0.5 ml HDB containing 7x10⁵ cpm (approximately 500 pM) ¹²⁵I activin A with or without 500 ng (37 nM) unlabeled activin A. Cells were 15 diluted with 1 ml HDB, pelleted by centrifugation and resuspended in 0.5 ml HDB. Disuccinimidyl suberate (DSS; freshly dissolved in DMSO) was added to 500 μM , and the cells incubated at 0° for 30 minutes. The cross-linking was terminated by addition of 1 ml 50 mM Tris-HCl pH 7.5, pelleted NaCl. cells were mMthen the 20 100 centrifugation, resuspended in 100 μ l 50 mM Tris-HCl pH 7.5, 1% Triton X-100 and incubated at 0° for 60 minutes. The samples were centrifuged 5 minutes at 13,000xg, and the Triton-soluble supernatants analyzed by SDS-PAGE using 8.5% 25 polyacrylamide gels. The gels were dried and subjected to autoradiography for 4-14 days.

EXAMPLE VII RNA Blot Analysis

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Total RNA was purified from tissue culture cells and tissues using LiCl precipitation. 20 μg total RNA was run on 1.2% agarose, 2.2M formaldehyde gels, blotted onto nylon membranes (Hybond - NEN), and hybridized with a 0.6 kb KpnI fragment (see Figure 3) which had been labeled with ³²P by random priming using reagents from US Biochemicals. Hybridization was performed at 42° in 50% formamide, and

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the filters were washed at 65° in 0.2X SSC.

EXAMPLE VIII Sequence Analysis

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Full length mouse activin receptor clone encodes a protein of 513 amino acids, with a 5' untranslated region of 70 bp and a 3' untranslated region of 951 bp. pmActR2 does not contain a poly A tail, although it does have a potential poladenylylation site at bp 2251. The insert in clone pmActR1 had an additional 551 bp of 5' untranslated sequence, was identical in the overlapping range, stopped at the 3' end at base 1132 of pmActR2. methionine codon (ATG), at bp 71, in pmActR2 is in a 15 favorable context for translation initiation [Kozak, M., Nucl. Acids Res. 15:8125-8148 (1987)], and is preceded by an in-frame stop codon. pmActR1 contains 3 additional ATGs in the 5' untranslated region; however, none of these is in an appropriate context for initiation, and all are followed 20 by in-frame stop codons. While this unusually long 5' leader sequence may have functional significance, it is clearly not necessary for proper expression, because pmActR2, which lacks most of that sequence, can be functionally expressed in COS cells (see below).

25

Hydropathy analysis using the method of Kyte and Doolittle [J. Mol. Biol. 157:105-132 (1982)] revealed two hydrophobic regions: a 10 amino acid stretch at the amino terminus assumed to be a single peptide, and a single putative 26 residue membrane-spanning region between amino acids 119-142 (see Figure 1 and Sequence ID No. 2). The signal peptide contains the conserved n-, h- and c- domains common to signal sequences; the site of cleavage of the signal peptide, before Ala¹, is predicted based on rules described by von Heijne [Biochim. Biophys. Act. 947:307-333 (1988)]. As is common for the cytoplasmic side of membrane-spanning domains, the predicted transmembrane

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region is closely followed by two basic amino acids. The mature mouse activin receptor is thus predicted to be a 494 amino acid type I membrane protein of Mr 54 kDa, with a 116 amino acid N-terminal extracellular ligand binding domain, and a 346 amino acid intracellular signalling domain.

Comparision of the activin receptor sequence to the sequence databases revealed structural similarity in the intracellular domain to a number of receptor and non-10 receptor kinases. Analysis of the sequences of all kinases has led to the identification of a 300 amino acid kinase domain characterized by 12 subdomains containing a number of highly conserved amino acids [Hanks, S.K. and Quinn, A.M., Meth. Enzymol. 200:38-62 (1991) and Hanks et al., 15 Science 241:42-52 (1988)]; the activin receptor sequence has all of these conserved subdomains in the proper order (Figure 4). A conserved Gly in subdomain I is replaced by Ala 180 in the activin receptor, but this residue has also been observed in other kinases. Based upon structural 20 relatedness, therefore, this receptor is expected to be a functional protein kinase.

The sequences in two of these subdomains (VIB and VIII) can be used to predict tyrosine vs. serine/threonine substrate specificity [Hanks et al., (1988) supra]. The sequence of the mouse activin receptor in both of these subdomains is characteristic of serine kinases.

Table 2 Kinase Domain Predictive Sequences

				!
Subdomain	VIB	SEQ ID NO.	VIII	SEO ID NO.
serine kinase consensus	DLKPEN	2	G(T/S)XX(Y/F)X	9
activin receptor	DIKSKN	7	GTRRYM	æ
tyrosine kinase consensus	DLAARN	O	XP(I/V)(K/R)W(T/M)	10

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Therefore, the activin receptor is expected to have serine/threonine specificity. Furthermore, the activin receptor does not have a tyrosine residue in the standard autophosphorylation region between subdomains VII 5 and VIII, indicating that it is not a standard tyrosine The receptor could potentially autophosphorylate kinase. at Ser 333 or Thr 337. One interesting additional possibility is that the activin receptor kinase may have specificity for serine, threonine and tyrosine residues. 10 kinases with these properties have recently been described [see, for example, Howell et al., Mol. Cell. Biol. 11:568-572 (1991), Stern et al., Mol. Cell. Biol. <u>11</u>:987-1001 (1991) and Featherstond, C. and Russell, P., Nature <u>349</u>:808-811 (1991)].

15

35

Phylogenetic analysis of the activin receptor compared to 161 other kinase sequences revealed that the activin receptor and the C.elegans protein, daf-1 [Georgi et al., Cell 61:635-645 (1990)] may constitute a separate 20 subfamily of kinases (see Figure 6). daf-1 is a putative transmembrane receptor involved in the developmental arrest of a non-feeding larval state and shares 32% identity with the activin receptor (see Figure 6). Like the activin receptor, daf-1 is predicted to be a transmembrane 25 serine/threonine-specific kinase; furthermore, both daf and the activin receptor have short, conserved inserts in the kinase domain sequence between subdomains VIA-VIB and X-XI that are not present in any other kinase (underlined in Figure 4B). This additional similarity lends credence to 30 their belonging to a unique subfamily of kinases. activin receptor is quite distantly related (18% amino acid sequence identity) to the only other known transmembrane serine/threonine protein kinase, encloded by the ZmPK gene of maize [Walker, J.C. and Zhang, R., Nature 345:743-746 (1990)].

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The extracellular domain of the activin receptor did not show similarity to any other sequences in the databases. This ligand binding domain is relatively small in comparison to those found in other growth factor receptors, but like those receptors this domain has a high cysteine content. The pattern of these Cys residues, however, is not like either an immunoglobulin fold or the cysteine rich repeats of the EGF receptor. There are also two potential sites of N-linked glycosylation in the extracellular domain, as well as a number of potential phosphorylation sites for protein kinase C and casein kinase II in the intracellular domain.

EXAMPLE IX

15 Binding Properties of the Cloned Activin Receptor

To verify that the cloned receptor is activin specific, competition binding experiments were performed on COS cells transiently transfected with either pmActR1 or 20 pmActR2. Cells transfected with either construct bound activin A with a single high affinity component (Kd = 180 pM; Figure 5), indicating that a (structurally complete) intracellular kinase domain is not required for ligand binding. This binding affinity is 25 consistent with that measured on other activin-responsive cell types [see, for example, Campen, C.A. and Vale, W., Biochem. Biophys. Res. Comm. 157:844-849 (1988); Hino et al., J. Biol. Chem. 264:10309-10314 (1989); Sugino et al., J. Biol. Chem. 263: 15249-15252 (1988); and Kondo et al., 30 Biochem. Biophys. Res. Comm, <u>161</u>:1267-1272 (1989)]. Untransfected COS cells do not bind activin A. transfected cultures as a whole expressed approximately 26,000 receptors per cell; however, because only 15% of the cells express the transfected gene (as measured by quantitating transfected cells as a fraction of all cells following dipping in emulsion), each transfected cell expressed an average of 175,000 receptors per cell. The

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level of expression per cell varies considerably, though, based on the number of accumulated silver grains. This value is comparable to the expression of other transfected cell surface proteins in COS cells.

5

Binding of iodinated activin A to COS cells transiently transfected with pmActR2 could be competed by activin B with slightly reduced potency compared to activin A; by inhibin A with approximately 10-fold lower potency; 10 and could not be competed by TGF-B1 (Figure 5B). affinity and specificity of binding match those observed following binding of activin A to a number of other activin-responsive cell types. Although activin B appears to bind the transfected receptor with lower affinity than 15 activin A, the activin B preparation used in these experiments may have suffered a reduction in potency, based on a comparison of bioactivity with activin A, since the recombinant synthesis of the activin B employed herein had been carried out some time ago [recombinant synthesis of 20 activin B is described by Mason et al., in Mol. Endocrinol. 3: 1352-1358 (1989)]. It is likely that this cDNA encodes a receptor for multiple forms of activin.

The size of the cloned activin receptor was

25 analyzed by affinity cross-linking 125I activin A to COS

cells transfected with pmActR2 using the bifunctional

chemical cross-linker, disuccinimidyl suberate (DSS). A

major cross-linked band of 84 kDa was observed in

transfected, but not in untransfected cells. Subtracting

30 the molecular weight of activin, this represents a protein

of 56 kDa, which corresponds well to the molecular weight

predicted from the nucleic acid sequence data. Cross
linking 125I activin A to AtT20 cells yields a major band of

65 kDa, with minor bands of approximately 78 and 84 kDa.

35 The size of the largest band matches that generated by the

cloned receptor. The smaller bands could be either

separate proteins, different phosphorylated forms of the

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same protein, or degradation products of the full length clone; the sequences DKKRR at amino acid 35 and KKKR at amino acid 416 could be potential sites of proteolysis. Alternatively, these bands could come from alternatively spliced products of the same gene.

The 84 and 65 kDa cross-linked bands have also been observed in other activin-responsive cell types [Hino, supra; Centrella et al., Mol. Cell. Biol. 11:250-258 (1991)], and interpreted to represent the signalling receptor, although complexes of other sizes have also been seen as well. The size of the activin receptor is very similar to a putative TGF-B receptor, to the limited extent it has been characterized by chemical cross-linking [see 15 Massague et al., Ann. N.Y. Acad. Sci. 593: 59-72 (1990)].

EXAMPLE X Expression of Activin Receptor mRNA

20 The distribution of activin receptor mRNA was analyzed by Northern blot. Two mRNA species, of 6.0 and 3.0 kb, were observed in AtT20 cells as well as a number of mouse tissues, including brain, testis, pancreas, liver and The total combined size of the inserts from kidney. 25 pmActR1 and pmActR2 is 3.1 kb, which corresponds to the size of the smaller transcript. Neither the extent of similarity between the two mRNAs, nor the significance of having two transcripts is clear. The genes for several other hormone receptors have been shown to be alternatively spliced to generate both a cell surface receptor and a 30 soluble binding protein, and it is possible that the activin receptor is processed in a similar manner.

Interestingly, the relative abundance of the two transcripts varies depending on the source. While AtT20 cells have approximately equal levels of both mRNAs, most tissues had much greater levels of the 6.0 kb transcript,

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with little or no expression of the 3.0 kb transcript. Testis, on the other hand, had a greater amount of the 3.0 kb band. Expression of activin receptor mRNA in brain, liver and testis is in accord with described biological actions of activin in those tissues [Mine et al., Endocrinol. 125:586-591 (1989); Vale et al., Peptide Growth Factors and Their Receptors, Handbook of Experimental Pharmacology, M.A. Sporn and A.B. Roberts, ed., Springer-Verlag (1990), in press].

10

EXAMPLE XI

Identification of a Human Activin Receptor

A human testis library (purchased from Clontech; 15 catalog no. HL1010b) was probed with the full length mouse activin receptor gene (see Sequence ID No. 1) under the following conditions:

Hybridization stringency:

20% formamide, 6X SSC at 42°C;

20 Wash stringency:

2X SSC, 0.1% SDS at 42°C.

A sequence which is highly homologous with the mouse activin receptor was identified (Sequence ID No. 1').

25 Due to the high degree of homology between this receptor and the mouse activin receptor, this receptor is designated as the human form of the activin receptor from the same subclass as the mouse receptor described above.

30

EXAMPLE XII

Identification of a Xenopus Activin Receptor

A Xenopus stage 17 embryo cDNA library (prepared as described by Kintner and Melton in Development 99: 311-35 325 (1987) was probed with the full length mouse activin receptor gene (see Sequence ID No. 1) under the following conditions:

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<u>Hybridization stringency:</u>

20% formamide, 6X SSC at 42°C;

Wash stringency:

2X SSC, 0.1% SDS at 42°C.

5

A sequence having a substantial degree of homology with respect to the mouse activin receptor was identified (Sequence ID No. 3). The degree of overall amino acid homology (relative to the mouse acitvin receptor) is only about 69% (with 77% homology in the intracellular domain and 58% homology in the extracellular domain). Due to the moderate degree of homology between this receptor and the mouse activin receptor, this receptor is designated as an activin receptor from a different subclass than the mouse receptor described above.

EXAMPLE XIII

Functional Assays of ActRs in Xenopus embryos

20 To determine whether xActRIIB can transmit a signal in response to activin, xActRIIB RNA was synthesized in vitro and injected into Xenopus embryos at two different concentrations. Injected embryos were allowed to develop to stage 9, at which time animal caps were dissected and 25 treated overnight with different concentrations of activin. The xActRIIB cDNA was cloned into rp64T [see Krieg and Melton in Methods in Enzymology, Abelson and Simon, Eds. (Academic Press, New York, 1987), vol. 155, p. 397] and transcribed in vitro to generate a capped, 30 xActRIIB RNA [see Melton et al., in Nucleic Acids Res. 12:7035 (1984) and Kintner in Neuron 1:545 (1988). Embryos at the two- to four-cell stage were injected with about 20 nl of RNA at concentrations of 0.02 ng/nl, or 0.1 ng/nl, spread between four quadrants of the animal pole. At stage 9, animal caps were removed from RNA-injected embryos and incubated in 0.5x modified mammalian Ringer's (MMR), 0.1% bovine serum albumin (BSA) with different

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concentrations of purified, porcine activin A (six caps per After 20 hours in culture, total RNA was incubation). prepared.

The response of the caps to activin was assessed quantifying muscle-specific RNA with actin ribonuclease protection assay as per Blackwell Weintraub, Science 250:1104 (1990). Embryos injected with 0.4 and 2.0 ng of xActRIIB RNA were approximately 10- and 10 100-fold more sensitive, respectively, to activin than control embryos. The low amount of muscle actin found in animal caps in the absence of added activin A is probably a consequence of contamination of the animal cap with a small amount of marginal zone tissue.

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The amount of muscle actin decreased with increasing concentration of activin in the embryos injected with 2 ng of xActRIIB RNA. This is consistent with the observation that isolated animal cap cells uniformly 20 exposed to different concentrations of activin only form muscle cells in response to a narrow range of activin concentrations [see Blackmann and Kadesch in Genes and Development 5:1057 (1990)]. The present results indicate that the concentration of ligand and the amount of receptor 25 are both important in determining the signal transmitted. Thus, the range of activin concentrations that lead to muscle differentiation is lower in animal cap cells from injected embryos, which are expressing more receptor than normal, than from uninjected embryos.

30

EXAMPLE XIV Analysis of kinase activity of mActRII

A fragment of cDNA corresponding to the entire 35 intracellular domain of mActRII (amino acids 143-494) was subcloned into the vector pGEX-2T [see Smith and Johnson in Gene 67:31-40 (1988)], creating a fusion protein between

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glutathione S-transferase (GST) and the putative kinase domain of the receptor. This plasmid was introduced into bacteria and the expressed fusion protein was purified using glutathione affinity chromatography as described by 5 Smith and Johnson. Approximately 100-200 ng of fusion protein, or of purified GST, were incubated with 25 μ Ci $[y^{-32}P]$ ATPin a buffer containing 50 mM Tris, 10 mM MgCl, for 30 minutes at 37°C. The products were analyzed by SDS-PAGE The fusion protein, but not the GST and autoradiography. 10 alone, became phosphorylated, indicating that the kinase domain of the fusion protein was functional. Phosphoamino acid analysis, performed according to Cooper et al. [Meth. Enzym. 99:387 (1983)], indicated that the predominant amino acid residue that became phosphorylated was threonine.

15

While the invention has been described in detail with reference to certain preferred embodiments thereof, it will be understood that modifications and variations are within the spirit and scope of that which is described and claimed.

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. SUMMARY OF SEQUENCES

Sequence ID No. 1 is the nucleic acid sequence (and the deduced amino acid sequence) of a cDNA encoding a mouse-derived activin receptor of the present invention.

Sequence ID No. 1' is a nucleic acid sequence encoding a human-derived activin receptor of the present invention. Sequence ID No. 1' is substantially the same as 10 Sequence ID No. 1, except that the codon for amino acid residue number 39 encodes lysine (i.e., nucleotides 185-187 are AAA or AAG), the codon for amino acid residue 92 encodes valine (i.e., nucleotides 344-346 are GTN, wherein N is A, C, G or T), and the codon for amino acid residue 15 number 288 encodes glutamine (i.e., nucleotides 932-934 are CAA or CAG).

Sequence ID No. 2 is the deduced amino acid sequence of a mouse-derived activin receptor of the present 20 invention.

Sequence ID No. 2' is an amino acid sequence for a human-derived activin receptor of the present invention. Sequence ID No. 2' is substantially the same as Sequence ID No. 2, except that amino acid residue number 39 is lysine, amino acid residue 92 is valine, and amino acid residue number 288 is glutamine.

Sequence ID No. 3 is the nucleic acid sequence 30 (and the deduced amino acid sequence) of a cDNA encoding a Xenopus-derived activin receptor of the present invention.

Sequence ID No. 4 is the deduced amino acid sequence of a Xenopus-derived activin receptor of the 35 present invention.

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Sequence ID No. 5 is the amino acid sequence of the VIB subdomain of the serine kinase consensus sequence.

Sequence ID No. 6 is the amino acid sequence of the VIII subdomain of the serine kinase consensus sequence.

Sequence ID No. 7 is the amino acid sequence of the VIB subdomain of the invention activin receptor.

Sequence ID No. 8 is the amino acid sequence of the VIII subdomain of the invention activin receptor.

Sequence ID No. 9 is the amino acid sequence of the VIB subdomain of the tyrosine kinase consensus 15 sequence.

Sequence ID No. 10 is the amino acid sequence of the VIII subdomain of the tyrosine kinase consensus sequence.

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SEQUENCE LISTING

(1) GENERAL	INFORMATION
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- (i) APPLICANT: Mathews, Ph.D., Lawrence S. Vale, Ph.D., Wylie W.
- (ii) TITLE OF INVENTION: CLONING AND RECOMBINANT PRODUCTION OF RECEPTOR(S) OF THE ACTIVIN/TGF-BETA SUPERFAMILY
- (iii) NUMBER OF SEQUENCES: 10
- (iv) CORRESPONDENCE ADDRESS:
 - (A) ADDRESSEE: PRETTY, SCHROEDER, BRUEGGEMANN & CLARK
 - (B) STREET: 444 South Flower Street, Suite 2000
 - (C) CITY: Los Angeles
 - (D) STATE: California
 - (E) COUNTRY: United States
 - (F) ZIP: 90071-2921
- (v) COMPUTER READABLE FORM:
 - (A) MEDIUM TYPE: Floppy disk
 - (B) COMPUTER: IBM PC compatible (C) OPERATING SYSTEM: PC-DOS/MS-DOS
 - (D) SOFTWARE: PatentIn Release #1.0, Version #1.25
- (vi) CURRENT APPLICATION DATA:
 - (A) APPLICATION NUMBER: US
 - (B) FILING DATE: 08-MAY-1992
 - (C) CLASSIFICATION:
- (viii) ATTORNEY/AGENT INFORMATION:
 - (A) NAME: Reiter, Mr., Stephen E.
 - (B) REGISTRATION NUMBER: 31192
 - (C) REFERENCE/DOCKET NUMBER: P31 9309/FP31 9291
 - (ix) TELECOMMUNICATION INFORMATION:
 - (A) TELEPHONE: (619) 546-4737
 - (B) TELEFAX: (619) 546-9392
- (2) INFORMATION FOR SEQ ID NO:1:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 2563 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: cDNA
 - (ix) FEATURE:
 - (A) NAME/KEY: CDS
 - (B) LOCATION: 71..1609
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:1:

CTCCGAGGAA GACCCAGGGA ACTGGATATC TAGCGAGAAC TTCCTACGGC TTCTCCGGCG

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CCTCGGGAAA ATG GGA GCT GCT GCA AAG TTG GCG TTC GCC GTC TTT CTT Met Gly Ala Ala Lys Leu Ala Phe Ala Val Phe Leu

5

		TGC Cys					Ile									:	157
	Leu	TTC Phe				Asn									ACT Thr 45	:	205
		GAA Glu			Tyr											:	253
GCT Ala	ACC	TGG Trp	AAG Lys 65	AAT Asn	ATT	TCT Ser	GGT Gly	TCC Ser 70	ATT	GAA Glu	ATA Ile	GTG Val	AAG Lys 75	CAA Gln	GGT Gly	3	301
		CTG Leu 80													GAA Glu	3	349
AAA Lys	AAA Lys 95	GAC Asp	AGC Ser	CCT Pro	GAA Glu	GTG Val 100	TAC Tyr	TTT Phe	TGT Cys	TGC Cys	TGT Cys 105	GAG Glu	GGC Gly	AAT Asn	ATG Met	3	397
		GAA Glu														4	145
ACT Thr	TCA Ser	AAT Asn	CCT Pro	GTT Val 130	ACA Thr	CCG Pro	AAG Lys	CCA Pro	CCC Pro 135	TAT Tyr	TAC Tyr	AAC Asn	ATT Ile	CTG Leu 140	CTG Leu	4	193
TAT Tyr	TCC Ser	TTG Leu	GTA Val 145	CCA Pro	CTA Leu	ATG Met	TTA Leu	ATT Ile 150	GCA Ala	GGA Gly	ATT Ile	GTC Val	ATT Ile 155	TGT Cys	GCA Ala	5	541
TTT Phe	TGG Trp	GTG Val 160	TAC Tyr	AGA Arg	CAT His	CAC His	AAG Lys 165	ATG Met	GCC Ala	TAC Tyr	CCT Pro	CCT Pro 170	GTA Val	CTT Leu	GTT Val	5	89
CCT Pro	ACT Thr 175	CAA Gln	GAC Asp	CCA Pro	GGA Gly	CCA Pro 180	CCC Pro	CCA Pro	CCT Pro	TCC Ser	CCA Pro 185	TTA Leu	CTA Leu	GGG Gly	TTG Leu	6	537
AAG Lys 190	CCA Pro	TTG Leu	CAG Gln	CTG Leu	TTA Leu 195	GAA Glu	GTG Val	AAA Lys	GCA Ala	AGG Arg 200	GGA Gly	AGA Arg	TTT Phe	GGT Gly	TGT Cys 205	6	85
GTC Val	TGG Trp	AAA Lys	GCC Ala	CAG Gln 210	TTG Leu	CTC Leu	AAT Asn	GAA Glu	TAT Tyr 215	GTG Val	GCT Ala	GTC Val	AAA Lys	ATA Ile 220	TTT Phe	7	'33
CCA Pro	ATA Ile	CAG Gln	GAC Asp 225	AAA Lys	CAG Gln	TCC Ser	TGG Trp	CAG Gln 230	AAT Asn	GAA Glu	TAT Tyr	GAA Glu	GTC Val 235	TAT Tyr	AGT Ser	7	81
CTA Leu	CCT Pro	GGA Gly 240	ATG Met	AAG Lys	CAT His	Glu	AAC Asn 245	ATA Ile	CTA Leu	CAG Gln	TTC Phe	ATT Ile 250	GGT Gly	GCA Ala	GAG Glu	8	29
AAA Lys	AGA Arg 255	GGC Gly	ACC Thr	AGT Ser	GTG Val	GAT Asp 260	GTG Val	GAC Asp	CTG Leu	TGG Trp	CTA Leu 265	ATC Ile	ACA Thr	GCA Ala	TTT Phe	8	77
CAT His 270	GAA Glu	AAG Lys	GGC Gly	TCA Ser	CTG Leu 275	TCA Ser	GAC Asp	TTT Phe	Leu	AAG Lys 280	GCT Ala	AAT Asn	GTG Val	GTC Val	TCT Ser 285	9	25

TGG Trp	AAT Asn	GAA Glu	CTT Leu	TGT Cys 290	CAT His	ATT Ile	GCA Ala	GAA Glu	ACC Thr 295	ATG Met	GCT Ala	AGA Arg	GGA Gly	TTG Leu 300	GCA Ala	973
TAT Tyr	TTA Leu	CAT His	GAG Glu 305	GAT Asp	ATA Ile	CCT Pro	GGC Gly	TTA Leu 310	AAA Lys	GAT Asp	GGC Gly	CAC His	AAG Lys 315	CCT Pro	GCA Ala	1021
ATC Ile	TCT Ser	CAC His 320	AGG Arg	GAC Asp	ATC Ile	AAA Lys	AGT Ser 325	AAA Lys	AAT Asn	GTG Val	CTG Leu	TTG Leu 330	AAA Lys	AAC Asn	AAT Asn	1069
CTG Leu	ACA Thr 335	Ala	TGC Cys	ATT Ile	GCT Ala	GAC Asp 340	TTT Phe	GGG Gly	TTG Leu	GCC Ala	TTA Leu 345	AAG Lys	TTC Phe	GAG Glu	GCT Ala	1117
GGC Gly 350	AAG Lys	TCT Ser	GCA Ala	GGT Gly	GAC Asp 355	ACC Thr	CAT His	GGG Gly	CAG Gln	GTT Val 360	GGT Gly	ACC Thr	CGG Arg	AGG Arg	TAT Tyr 365	1165
ATG Met	GCT Ala	CCA Pro	GAG Glu	GTG Val 370	TTG Leu	GAG Glu	GGT Gly	GCT Ala	ATA Ile 375	AAC Asn	TTC Phe	CAA Gln	AGG Arg	GAC Asp 380	GCA Ala	1213
TTT Phe	CTG Leu	AGG Arg	ATA Ile 385	GAT Asp	ATG Met	TAC Tyr	GCC Ala	ATG Met 390	GGA Gly	TTA Leu	GTC Val	CTA Leu	TGG Trp 395	GAA Glu	TTG Leu	1261
GCT Ala	TCT Ser	CGT Arg 400	TGC Cys	ACT Thr	GCT Ala	GCA Ala	GAT Asp 405	GGA Gly	CCC Pro	GTA Val	GAT Asp	GAG Glu 410	TAC Tyr	ATG Met	TTA Leu	1309
CCA Pro	TTT Phe 415	GAG Glu	GAA Glu	GAA Glu	ATT Ile	GGC Gly 420	CAG Gln	CAT His	CCA Pro	TCT Ser	CTT Leu 425	GAA Glu	GAT Asp	ATG Met	CAG Gln	1357
GAA Glu 430	GTT Val	GTT Val	GTG Val	CAT His	AAA Lys 435	AAA Lys	AAG Lys	AGG Arg	CCT Pro	GTT Val 440	TTA Leu	AGA Arg	GAT Asp	TAT Tyr	TGG Trp 445	1405
CAG Gln	AAA Lys	CAT His	GCA Ala	GGA Gly 450	ATG Met	GCA Ala	ATG Met	CTC Leu	TGT Cys 455	GAA Glu	ACG Thr	ATA Ile	GAA Glu	GAA Glu 460	TGT Cys	1453
TGG Trp	GAT Asp	CAT His	GAT Asp 465	GCA Ala	GAA Glu	GCC Ala	AGG Arg	TTA Leu 470	TCA Ser	GCT Ala	GGA Gly	TGT Cys	GTA Val 475	GGT Gly	GAA Glu	1501
AGA Arg	ATT Ile	ACT Thr 480	CAG Gln	ATG Met	CAA Gln	AGA Arg	CTA Leu 485	ACA Thr	TAA neA	ATC Ile	ATT Ile	ACT Thr 490	ACA Thr	GAG Glu	GAC Asp	1549
ATT Ile	GTA Val 495	ACA Thr	GTG Val	GTC Val	ACA Thr	ATG Met 500	GTG Val	ACA Thr	TAA neA	GTT Val	GAC Asp 505	TTT Phe	CCT Pro	CCC Pro	aaa Lys	1597
GAA Glu 510			CTA Leu	TGAT	ggtg	GC A	CCG1	CTGI	'A CA	CACT	GAGG	ACT	GGGA	CTC		1649
TGAA	CTGG	AG C	TGCT	'AAGC	TAA	.GGAA	AGTG	CTI	'AGTI	GAT	TTTC	TGTG	TG · A	AATG	AGTA	3 1709
GATG	CCTC	CA G	GACA	TGTA	C GC	AAGC	AGCC	CCI	TGTG	GAA	AGCA	TGGA	TC I	GGGA	GATG	3 1769
ATCT	GGGA	AA C	TTAC	TGCA	T CG	TCTG	CAGO	: ACA	.GATA	TGA	AGAG	GAGI	CT A	AGGG	AAAA	G 1829
CTGC	AAAC	TG I	'AAAG	AACI	T CI	GAAA	ATGI	ACT	CGAA	GAA	TGTG	GCCC	TC I	CCAA	atcai	A 1889

GGATCTTTTC	GACCTGGCTA	ATCAAGTATT	TGCAAAACTG	ACATCAGATT	TCTTAATGTC	1949
TGTCAGAAGA	CACTAATTCC	TTAAATGAAC	TACTGCTATT	TTTTTTAAAT	GAAAAACTTT	2009
TCATTTCAGA	TTTTAAAAAG	GGTAACTTTT	TATTGCATTT	GCTGTTGTTT	CTATAAATGA	2069
CTATTGTAAT	GCCAACATGA	CACAGCTTGT	GAATGTGTAG	TGTGCTGCTG	TTCTGTGTAC	2129
ATAGTCATCA	AAGTGGGGTA	CAGTAAAGAG	GCTTCCAAGC	ATTACTTTAA	CCTCCCTCAA	2189
CAAGGTATAC	CTCAGTTCCA	CGGTTGTTAA	ATTATAAAAT	TGAAAACACT	AACAGAATTT	2249
GAATAAATCA	GTCCATGTTT	TATAACAAGG	TTAATTACAA	ATTCACTGTG	TTATTTAAGA	2309
AAAAATGGTA	AGCTATGCTT	AGTGCCAATA	GTAAGTGGCT	ATTTGTAAAG	CAGTGTTTTA	2369
GCTTTTCTTC	TACTGGCTTG	TAATTTAGGG	AAAACAAGTG	CTGTCTTTGA	AATGGAAAAG	2429
AATATGGTGT	CACCCTACCC	CCCATACTTA	TATCAAGGTC	CCAAAATATT	CTTTTCCATT	2489
TCAAAGACAG	CACTTTGAAA	ACCCTAAATT	ACAAGCCAGT	AGAAGAAAAG	CTAAAACACG	2549
CTTTACAAAT	AGCC					2563

(2) INFORMATION FOR SEQ ID NO:2:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 513 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:

Met Gly Ala Ala Ala Lys Leu Ala Phe Ala Val Phe Leu Ile Ser Cys
15

Ser Ser Gly Ala Ile Leu Gly Arg Ser Glu Thr Gln Glu Cys Leu Phe
20

Phe Asn Ala Asn Trp Glu Arg Asp Arg Thr Asn Gln Thr Gly Val Glu
35

Pro Cys Tyr Gly Asp Lys Asp Lys Arg Arg His Cys Phe Ala Thr Trp
50

Lys Asn Ile Ser Gly Ser Ile Glu Ile Val Lys Gln Gly Cys Trp Leu 65 70 75 80

Asp Asp Ile Asn Cys Tyr Asp Arg Thr Asp Cys Ile Glu Lys Lys Asp 85 90 95

Ser Pro Glu Val Tyr Phe Cys Cys Cys Glu Gly Asn Met Cys Asn Glu 100 105 110

Lys Phe Ser Tyr Phe Pro Glu Met Glu Val Thr Gln Pro Thr Ser Asn 115 120 125

Pro Val Thr Pro Lys Pro Pro Tyr Tyr Asn Ile Leu Leu Tyr Ser Leu 130 135 140

Val Pro Leu Met Leu Ile Ala Gly Ile Val Ile Cys Ala Phe Trp Val 145 150 155 160

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Tyr Arg His His Lys Met Ala Tyr Pro Pro Val Leu Val Pro Thr Gln Asp Pro Gly Pro Pro Pro Pro Ser Pro Leu Leu Gly Leu Lys Pro Leu 185 Gln Leu Leu Glu Val Lys Ala Arg Gly Arg Phe Gly Cys Val Trp Lys Ala Gln Leu Leu Asn Glu Tyr Val Ala Val Lys Ile Phe Pro Ile Gln 210 215 220 Asp Lys Gln Ser Trp Gln Asn Glu Tyr Glu Val Tyr Ser Leu Pro Gly Met Lys His Glu Asn Ile Leu Gln Phe Ile Gly Ala Glu Lys Arg Gly Thr Ser Val Asp Val Asp Leu Trp Leu Ile Thr Ala Phe His Glu Lys 265 Gly Ser Leu Ser Asp Phe Leu Lys Ala Asn Val Val Ser Trp Asn Glu Leu Cys His Ile Ala Glu Thr Met Ala Arg Gly Leu Ala Tyr Leu His Glu Asp Ile Pro Gly Leu Lys Asp Gly His Lys Pro Ala Ile Ser His Arg Asp Ile Lys Ser Lys Asn Val Leu Leu Lys Asn Asn Leu Thr Ala Cys Ile Ala Asp Phe Gly Leu Ala Leu Lys Phe Glu Ala Gly Lys Ser Ala Gly Asp Thr His Gly Gln Val Gly Thr Arg Arg Tyr Met Ala Pro Glu Val Leu Glu Gly Ala Ile Asn Phe Gln Arg Asp Ala Phe Leu Arg Ile Asp Met Tyr Ala Met Gly Leu Val Leu Trp Glu Leu Ala Ser Arg 390 Cys Thr Ala Ala Asp Gly Pro Val Asp Glu Tyr Met Leu Pro Phe Glu Glu Glu Ile Gly Gln His Pro Ser Leu Glu Asp Het Gln Glu Val Val 425 Val His Lys Lys Lys Arg Pro Val Leu Arg Asp Tyr Trp Gln Lys His Ala Gly Met Ala Met Leu Cys Glu Thr Ile Glu Glu Cys Trp Asp His Asp Ala Glu Ala Arg Leu Ser Ala Gly Cys Val Gly Glu Arg Ile Thr Gln Met Gln Arg Leu Thr Asn Ile Ile Thr Thr Glu Asp Ile Val Thr 490 Val Val Thr Met Val Thr Asn Val Asp Phe Pro Pro Lys Glu Ser Ser

(2) INFORMATION FOR SEC	ID	NO: 3:
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- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 2335 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(vii) IMMEDIATE SOURCE: (B) CLONE: XACTR

(ix) FEATURE:

- (A) NAME/KEY: CDS (B) LOCATION: 468..1997

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:3:

CCGCCCACAC AG	STGCAGTGA ATAATAGO	CC GTGCGGCCCC	TCCCCTCTTT CCCT	GGCAGT 60
TGTGTATCTG TC	CACATTGAA GTTTGGGC	TC CTGTGAGTCT	GAGCCTCCCC CTGT	GTCTCA 120
TGTGAAGCTG CT	GCTGCAGA AGGTGGAG	TC GTTGCATGAG	GGTGGGGGGA GTCG	CTGCTG 180
TTTGATCTGC CT	CTGCTCCC CATTCACA	CT CTCATTTCAT	TCCCACGGAT CCAC	ATTACA 240
ACTCGCCTTT AA	ACCCTTTCC CTGGCGGA	GC CCACGCGTCT	TTCATCCCTC CTGC	CGCGGC 300
CGCTGAGCGA CC	AGAGCGCG ACATTGTT	GC GGCGGGGAT	TGGGCGACAT TGTT	GCGAAT 360
AATCGGAGCT GC	TGGGGGG AACTGATA	CA ACGTTGCGAC	TGTAAAGGAA TTAA	CTCGGC 420
CGAATGGGAT TT	TATCTGTG TCGGTGAG	AG AAGCGGATCC	CAGGAGC ATG GGG Met Gly 1	
TCT GTA GCG C Ser Val Ala L 5	TG ACT TTT CTA CT eu Thr Phe Leu Le 10	T CTT CTT GCA u Leu Leu Ala	ACT TTC CGC GCA Thr Phe Arg Ala 15	GGC 524 Gly
TCA GGA CAC G Ser Gly His A 20	AT GAA GTG GAG AC sp Glu Val Glu Th 25	A AGA GAG TGC . r Arg Glu Cys 30	ATC TAT TAC AAT lle Tyr Tyr Asn	GCC 572 Ala 35
AAC TGG GAA C Asn Trp Glu L	TG GAG AAG ACC AA eu Glu Lys Thr As: 40	C CAA AGT GGG n Gln Ser Gly 45	GTG GAA AGC TGC Val Glu Ser Cys 50	GAA 620 Glu
Gly Glu Lys A	AC AAG CGA CTC CA sp Lys Arg Leu Hi 55	C TGT TAC GCG ! B Cys Tyr Ala ! 60	TCT TGG AGG AAC Ser Trp Arg Asn 65	AAT 668 Asn
TCG GGC TTC AS Ser Gly Phe II	TA GAG CTG GTG AA le Glu Leu Val Lyd 7:	s Lys Gly Cys !	TGG CTG GAT GAC Trp Leu Asp Asp 80	TTC 716 Phe
AAC TGT TAT GA ABN Cys Tyr As 85	AC AGA CAG GAA TG BP Arg Gln Glu Cy 90	T ATT GCC AAG (B Ile Ala Lyb (GAA GAA AAC CCC Glu Glu Amn Pro 95	CAA 764 Gln

GTC Val 100	TTT	TTC Phe	TGC Cys	TGC Cys	TGC Cys 105	GAG Glu	GGA Gly	AAC Asn	TAC Tyr	TGC Cys 110	AAC Asn	AAG Lys	AAA Lys	TTT Phe	ACT Thr 115	812
CAT His	TTG Leu	CCT Pro	GAA Glu	GTC Val 120	Glu	ACA Thr	TTT Phe	GAT Asp	CCG Pro 125	AAG Lys	CCC Pro	CAG Gln	CCG Pro	TCA Ser 130	GCC Ala	860
TCC Ser	GTA Val	CTG Leu	AAC Asn 135	ATT Ile	CTG Leu	ATC Ile	TAT Tyr	TCC Ser 140	CTG Leu	CTT Leu	CCA Pro	ATT Ile	GTT Val 145	GGT Gly	CTT Leu	908
TCC Ser	ATG Met	GCA Ala 150	Ile	CTC Leu	CTG Leu	GCG Ala	TTC Phe 155	TGG Trp	ATG Met	TAC Tyr	CGT Arg	CAT His 160	CGA Arg	AAG Lys	CCT Pro	956
CCC Pro	TAC Tyr 165	Gly	CAT His	GTA Val	GAG Glu	ATC Ile 170	AAT Asn	GAG Glu	GAC Asp	CCC Pro	GGT Gly 175	CTG Leu	CCC Pro	CCT Pro	CCA Pro	1004
TCT Ser 180	CCT Pro	CTG Leu	GTC Val	GGG Gly	CTG Leu 185	AAG Lys	CCG Pro	CTG Leu	CAG Gln	TTG Leu 190	CTG Leu	GAG Glu	ATA Ile	AAG Lys	GCG Ala 195	1052
CGA Arg	GGC Gly	CGT Arg	TTC Phe	GGT Gly 200	TGC Cys	GTC Val	TGG Trp	AAA Lys	GCT Ala 205	CGT Arg	CTG Leu	CTG	AAT Asn	GAA Glu 210	TAT Tyr	1100
GTC Val	GCA Ala	GTG Val	AAA Lys 215	ATC Ile	TTC Phe	CCC Pro	GTG Val	CAG Gln 220	GAT Asp	AAG Lys	CAG Gln	TCG Ser	TGG Trp 225	CAG Gln	TGT Cys	1148
GAG Glu	AAA Lys	GAG Glu 230	ATC Ile	TTC Phe	ACC Thr	ACG Thr	CCG Pro 235	GGC Gly	ATG Met	AAA Lys	CAT His	GAA Glu 240	AAC Asn	CTA Leu	TTG Leu	1196
GAG Glu	TTC Phe 245	ATT Ile	GCC Ala	GCT Ala	GAG Glu	AAG Lys 250	AGG Arg	GGA Gly	AGC Ser	AAC Asn	CTG Leu 255	GAG Glu	ATG Met	GAG Glu	CTG Leu	1244
TGG Trp 260	CTC Leu	ATC Ile	ACT Thr	GCA Ala	TTT Phe 265	CAT His	GAT Asp	AAG Lys	GGT Gly	TCT Ser 270	CTG Leu	ACG Thr	gac Asp	TAC Tyr	CTG Leu 275	1292
AAA Lys	GGG Gly	AAC Asn	TTG Leu	GTG Val 280	AGC Ser	TGG Trp	AAT Asn	GAA Glu	CTG Leu 285	TGT Cys	CAC His	ATA Ile	ACA Thr	GAA Glu 290	ACA Thr	1340
ATG Met	GCT Ala	CGT Arg	GGG Gly 295	CTG Leu	GCC Ala	TAC Tyr	TTA Leu	CAT His 300	GAA Glu	GAT Asp	GTG Val	CCC Pro	CGC Arg 305	TGT Cys	AAA Lys	1388
GGT Gly	GAA Glu	GGG Gly 310	CAC His	AAA Lys	CCT Pro	GCA Ala	ATC Ile 315	GCT Ala	CAC His	AGA Arg	GAT Asp	TTT Phe 320	AAA Lys	AGT Ser	AAG Lys	1436
AAT Asn	GTA Val 325	TTG Leu	CTA Leu	AGA Arg	AAC Asn	GAC Asp 330	CTG Leu	ACT Thr	GCG Ala	ATA Ile	TTA Leu 335	GCA Ala	GAC Asp	TTC Phe	GGG Gly	1484
CTG Leu 340	GCC Ala	GTA Val	CGA Arg	TTT Phe	GAG Glu 345	CCT Pro	GGA Gly	AAA Lys	CCT Pro	CCG Pro 350	GGA Gly	GAT Asp	ACA Thr	CAC His	GGG Gly 355	1532
CAG Gln	GTT Val	GGC Gly	ACC Thr	AGG Arg 360	AGG Arg	TAT Tyr	ATG Met	GCT Ala	CCT Pro 365	GAG Glu	GTT Val	CTA Leu	GAG Glu	GGA Gly 370	GCA Ala	1580

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											GAT Asp					:	1628
											ACA Thr					3	1676
											GAG Glu 415					1	1724
CCT Pro 420	TCC Ser	CTA Leu	GAG Glu	GAT Asp	CTG Leu 425	CAA Gln	GAA Glu	GTT Val	GTC Val	GTT Val 430	CAC His	AAG Lys	AAG Lys	ATA Ile	CGC Arg 435	1	1772
											GGT Gly					1	1820
TGC Cyb	GTC Val	ACC Thr	ATT Ile 455	GAA Glu	GAA Glu	TGC Cys	TGG Trp	GAC Asp 460	CAT His	GAT Asp	GCG Ala	GAA Glu	GCA Ala 465	CGG Arg	CTT Leu	1	1868
TCG Ser	GCA Ala	GGC Gly 470	TGC Cys	GTA Val	GAG Glu	GAG Glu	CGT Arg 475	ATT Ile	TCC Ser	CAA Gln	ATC Ile	CGT Arg 480	AAA Lys	TCA Ser	GTG Val	1	916
											GTT Val 495					1	.964
AAT Asn 500											TGAG	GTTI	CT I	TGGI	CTTTC	2	017
CAGA	CTCA	GT G	ACTI	TTAP	AA AA	AAAA	ACTO	ACG	AATG	CAG	CTGC	TATI	TT A	TCTI	GACTT	2	077
TTTA	TATA	TT T	TTTI	CTTG	G AT	'TTTA	CTTG	GAT	CGGA	TCA	ATTT	ACCA	GC A	CGTC	ATTCG	2	137
AAAG	TATT	A AA'	AAAA	AAAA	A CA	AAAC	AAAA	AAG	CAAA	AAC	AGAC	ATCT	CA G	CAAG	CATTC	2	197
AGGT	GCCG	AC T	TATG	AATG	C CA	ATAG	GTGC	AGG	AACT	TCA	GAAC	CTCA	AC A	AACI	CATTT	2	257
CTAG	AGAA	TG T	TCTC	CTGG	T TT	CCTT	TATC	TCA	GAAG	AGG	ACCC	ATAG	GA A	AACA	CCTAA	2	317
GTCA	AGCA	AA T	GCTG	CAG												2	335

(2) INFORMATION FOR SEQ ID NO:4:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 510 amino acids (B) TYPE: amino acid

 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:

Met Gly Ala Ser Val Ala Leu Thr Phe Leu Leu Leu Leu Ala Thr Phe

Tyr Asn Ala Asn Trp Glu Leu Glu Lys Thr Asn Gln Ser Gly Val Glu Ser Cys Glu Gly Glu Lys Asp Lys Arg Leu His Cys Tyr Ala Ser Trp 50 55 60 Arg Asn Asn Ser Gly Phe Ile Glu Leu Val Lys Lys Gly Cys Trp Leu 65 70 75 80 Asp Asp Phe Asn Cys Tyr Asp Arg Gln Glu Cys Ile Ala Lys Glu Glu Asn Pro Gln Val Phe Phe Cys Cys Cys Glu Gly Asn Tyr Cys Asn Lys 100 105 110 Lys Phe Thr His Leu Pro Glu Val Glu Thr Phe Asp Pro Lys Pro Gln Pro Ser Ala Ser Val Leu Asn Ile Leu Ile Tyr Ser Leu Leu Pro Ile Val Gly Leu Ser Met Ala Ile Leu Leu Ala Phe Trp Met Tyr Arg His Arg Lys Pro Pro Tyr Gly His Val Glu Ile Asn Glu Asp Pro Gly Leu 170 Pro Pro Pro Ser Pro Leu Val Gly Leu Lys Pro Leu Gln Leu Leu Glu Ile Lys Ala Arg Gly Arg Phe Gly Cys Val Trp Lys Ala Arg Leu Leu 195 200 205 Asn Glu Tyr Val Ala Val Lys Ile Phe Pro Val Gln Asp Lys Gln Ser Trp Gln Cys Glu Lys Glu Ile Phe Thr Thr Pro Gly Met Lys His Glu Asn Leu Leu Glu Phe Ile Ala Ala Glu Lys Arg Gly Ser Asn Leu Glu Met Glu Leu Trp Leu Ile Thr Ala Phe His Asp Lys Gly Ser Leu Thr Asp Tyr Leu Lys Gly Asn Leu Val Ser Trp Asn Glu Leu Cys His Ile Thr Glu Thr Met Ala Arg Gly Leu Ala Tyr Leu His Glu Asp Val Pro Arg Cys Lys Gly Glu Gly His Lys Pro Ala Ile Ala His Arg Asp Phe Lys Ser Lys Asn Val Leu Leu Arg Asn Asp Leu Thr Ala Ile Leu Ala Asp Phe Gly Leu Ala Val Arg Phe Glu Pro Gly Lys Pro Pro Gly Asp 345 Thr His Gly Gln Val Gly Thr Arg Arg Tyr Met Ala Pro Glu Val Leu Glu Gly Ala Ile Asn Phe Gln Arg Asp Ser Phe Leu Arg Ile Asp Met 370 380

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Tyr Ala Met Gly Leu Val Leu Trp Glu Ile Val Ser Arg Cys Thr Ala 385 390 395 400

Ala Asp Gly Pro Val Asp Glu Tyr Leu Leu Pro Phe Glu Glu Glu Ile 405 410 415

Gly Gln His Pro Ser Leu Glu Asp Leu Gln Glu Val Val His Lys 420 425 430

Lys Ile Arg Pro Val Phe Lys Asp His Trp Leu Lys His Pro Gly Leu 435 440 445

Ala Gln Leu Cys Val Thr Ile Glu Glu Cys Trp Asp His Asp Ala Glu 450 455 460

Ala Arg Leu Ser Ala Gly Cys Val Glu Glu Arg Ile Ser Gln Ile Arg 465 470 475

Lys Ser Val Asn Gly Thr Thr Ser Asp Cys Leu Val Ser Ile Val Thr 485 490 495

Ser Val Thr Asn Val Asp Leu Pro Pro Lys Glu Ser Ser Ile 500 505 510

SEQ ID NO.: 5

DLKPEN

SEQ ID No.: 6

G(T/S)XX(Y/F)X

SEQ ID NO.: 7

DIKSKN

SEQ ID NO.: 8

GTRRYM

SEQ ID NO.: 9

DLAARN

SEQ ID NO.: 10

XP(I/V)(K/R)W(T/M)

That which is claimed is:

1. A novel receptor protein characterized by having the following domains, reading from the N-terminal end of said protein:

an extracellular, ligand-binding domain, a hydrophobic, trans-membrane domain, and an intracellular, receptor domain having serine kinase-like activity.

- A protein according to Claim 1, further
 comprising a second hydrophobic domain at the amino terminus thereof.
- 3. A protein according to Claim 1, wherein said protein is further characterized by having sufficient binding affinity for at least one member of the activin/TGF-B superfamily of polypeptide growth factors such that concentrations of ≤ 10 nM of said polypeptide growth factor occupy ≥ 50% of the binding sites of said receptor protein.

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4. A protein according to Claim 3, wherein said protein is further characterized by:

having a greater binding affinity for activins than for inhibins,

25 having substantially no binding affinity for transforming growth factors-B, and

having substantially no binding affinity for non-activin-like proteins.

5. A protein according to Claim 1 having an amino acid sequence substantially the same as set forth in Sequence ID No. 2, Sequence ID No. 2', or Sequence ID No. 4.

6. A soluble, extracellular, ligand-binding protein, further characterized by:

having a sufficient binding affinity for at least one member of the activin/TGF-B superfamily of polypeptide growth factors such that concentrations of ≤ 10 nM of said polypeptide growth factor occupy ≥ 50% of the binding sites on said receptor protein, and

having at least about 30% sequence identity with respect to:

the sequence of amino acids 20-134 set forth in Sequence ID No. 2;

15

35

the sequence of amino acids 20-134 set forth in Sequence ID No. 2, wherein the arginine residue at position number 39 is replaced by a lysine, and the isoleucine at residue number 92 is replaced by a valine; or

the sequence of amino acids 21-132 set forth in Sequence ID No. 4.

7. A protein according to Claim 6, further characterized by:

having a greater binding affinity for activins than for inhibins,

having substantially no binding affinity for 25 transforming growth factors-B, and

having substantially no binding affinity for non-activin-like proteins.

- 8. A protein according to Claim 6 wherein said 30 protein comprises in the range of about 114-118 amino acids.
 - 9. A DNA encoding a mature protein according to Claim 1.
 - 10. A DNA encoding a mature protein according to Claim 3.

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- 11. A DNA encoding a precursor-form of the protein of Claim 1.
- 12. A DNA encoding a protein according to 5 Claim 2.
 - 13. A DNA encoding a soluble protein according to Claim 6.
- 10 14. A DNA encoding a soluble protein according to Claim 8.

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- 15. A DNA encoding a precursor-form of the protein of Claim 6.
- 16. A DNA according to Claim 9 having a contiguous nucleotide sequence substantially the same as: nucleotides 128 1609 of Sequence ID No. 1;

variations of nucleotides 128 - 1609 of Sequence

10 No. 1, wherein the codon for residue number 39 of
the encoded amino acid codes for lysine, the codon for
residue number 92 of the encoded amino acid codes for
valine, and the codon for residue number 288 of the
encoded amino acid encodes glutamine;

- nucleotides 528 1997 of Sequence ID No. 3; or variations of any of the above sequences which encode the same amino acid sequences, but employ different codons for some of the amino acids.
- 30 17. A DNA according to Claim 9 having a contiguous nucleotide sequence substantially the same as: nucleotides 71 1609 of Sequence ID No. 1;

variations of nucleotides 71 - 1609 of Sequence ID No. 1, wherein the codon for residue number 39 of the encoded amino acid codes for lysine, the codon for residue number 92 of the encoded amino acid codes for valine, and the codon for residue number 288 of the

encoded amino acid encodes glutamine;

nucleotides 468 - 1997 of Sequence ID No. 3; or variations of any of the above sequences which encode the same amino acid sequences, but employ different codons for some of the amino acids.

- 18. A DNA according to Claim 9 having a contiguous nucleotide sequence substantially the same as set forth in Sequence ID No. 1, Sequence ID No. 1' or 10 Sequence ID No. 3.
- 19. A DNA according to Claim 13 having a contiguous nucleotide sequence substantially the same as nucleotides 71 127 of Sequence ID No. 1, or nucleotides 15 468-527 of Sequence ID No. 3.
 - 20. A method for the recombinant production of activin receptor(s), said method comprising expressing the DNA of Claim 9 in a suitable host cell.

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21. A method for the recombinant production of soluble activin receptor(s), said method comprising expressing the DNA of Claim 13 in a suitable host cell.

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- 22. A DNA fragment useful as a hybridization probe, wherein said DNA fragment comprises at least a portion of the DNA according to Claim 9, and wherein said DNA fragment is labeled with a readily detectable substituent.
- 23. A DNA fragment according to Claim 22 wherein said readily detectable substituent is selected from a radiolabeled molecule, a fluorescent molecule, an enzyme, or a ligand.

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24. A method to identify clones encoding receptors of the activin/TGF- β superfamily, said method comprising:

screening a genomic or cDNA library with a DNA fragment according to Claim 22 under low stringency hybridization conditions, and

identifying those clones which display a substantial degree of hybridization to said DNA fragment.

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- 25. A method for screening a collection of compounds to determine those compounds which bind to receptors of the activin/TGF-B superfamily, said method comprising employing the receptor of claim 1 in a competitive binding assay.
- 26. A bioassay for evaluating whether compounds are agonists for receptor protein(s) according to Claim 1,20 or functional modified forms of said receptor protein(s), said bioassay comprising:
 - (a) culturing cells containing:

DNA which expresses said receptor protein(s) or functional modified forms of said receptor protein(s), and

DNA encoding a hormone response element operatively linked to a reporter gene,

wherein said culturing is carried out in the presence of at least one compound whose ability to induce transcription activation activity of said receptor protein is sought to be determined; and thereafter

(b) monitoring said cells for expression of said reporter gene.

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27. A bioassay for evaluating whether compounds are antagonists for receptor protein(s) according to Claim 1, or functional modified forms of said receptor protein(s), said bioassay comprising:

5 (a) culturing cells containing:

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DNA which expresses said receptor protein(s), or functional modified forms of said receptor protein(s), and

DNA encoding a hormone response element operatively linked to a reporter gene;

wherein said culturing is carried out in the presence of:

increasing concentrations of at least one compound whose ability to inhibit transcription activation of said receptor protein(s) is sought to be determined, and

- a fixed concentration of at least one agonist for said receptor protein(s), or functional modified forms of said receptor protein(s); and thereafter
- (b) monitoring in said cells the level of expression of the product of said reporter gene as a function of the concentration of said compound, thereby indicating the ability of said compound to inhibit activation of transcription.
- 28. A method for modulating the transcription trans-activation of activin receptor(s), said method comprising:
- contacting said receptor with an effective, modulating amount of the protein of Claim 6.
 - 29. An antibody generated against the protein of Claim 6.
 - 30. An antibody according to Claim 29, wherein said antibody is a monoclonal antibody.

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31. A method for modulating the transcription trans-activation of activin receptor(s), said method comprising:

contacting said receptor with a modulating, effective amount of the antibody of Claim 29.

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C-terminus ------ Intracellular domain -------- kinase domain ---Figure 1 Trans-membrane domain Ligand-binding domain Second hydrophobic domain N-terminus

SUBSTITUTE SHEET

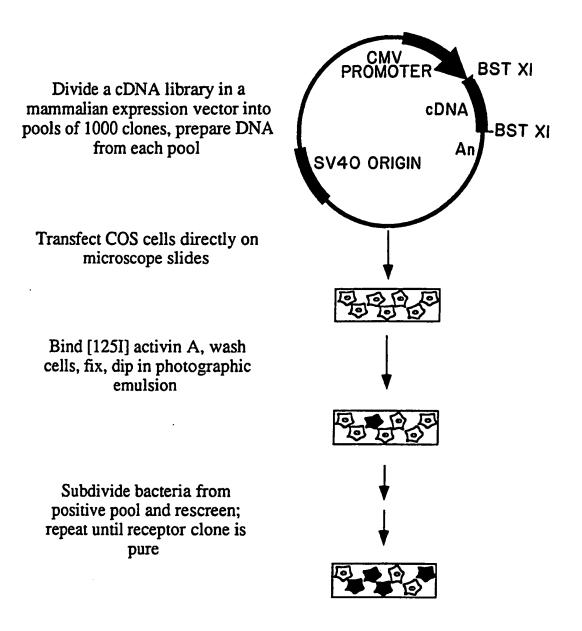
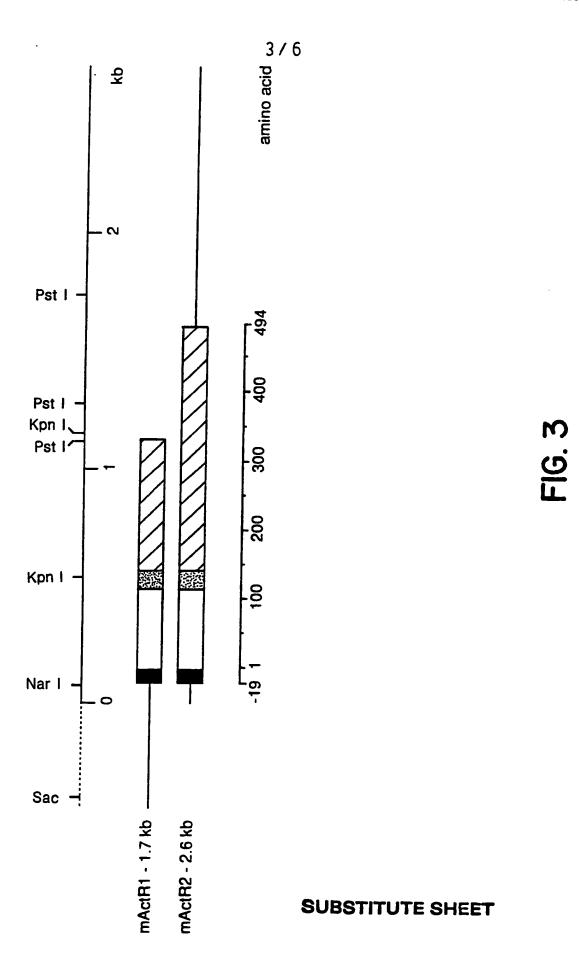


FIG. 2



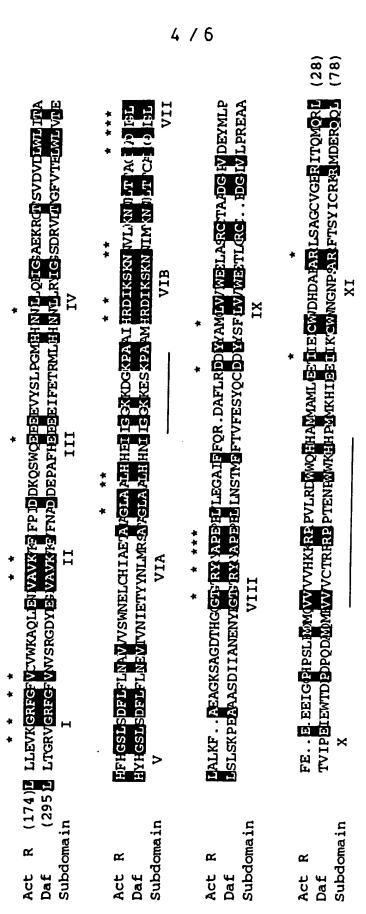
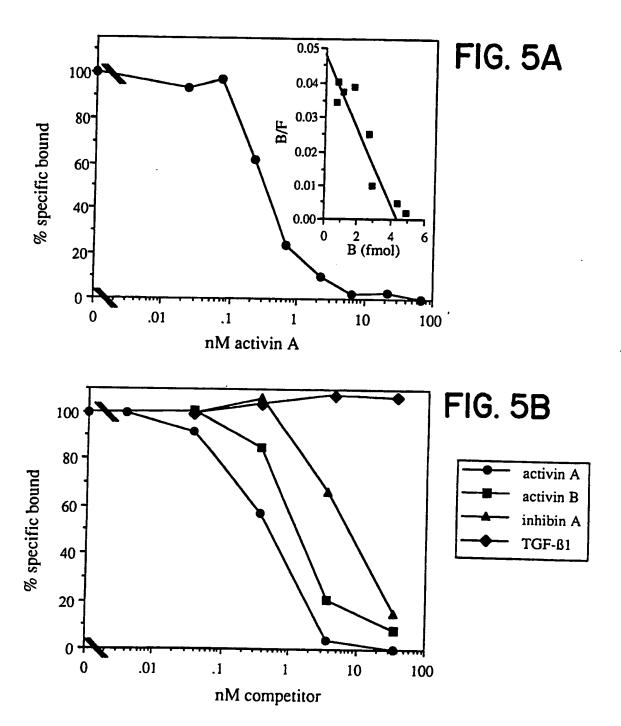
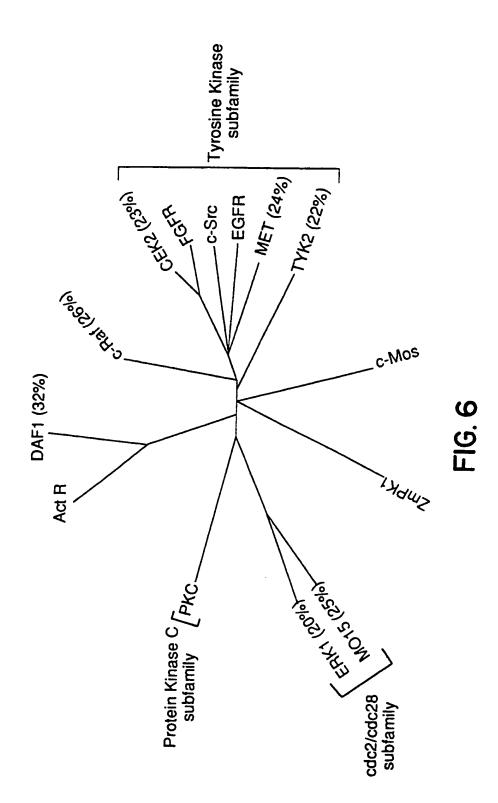


FIG. 4





SUBSTITUTE SHEET

International Application No

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		t Classification (IPC) or to both National		
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	C12N15/6	2; A61K37/02;	C12F21/U0,	W01V33\ 333
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		GO1N; A61K		
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Category °	Citation of De	ocument, 11 with Indication, where approp	priate, of the relevant passages 44	Relevant to Claim No.13
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X	CELL	, no. 3, 18 May 1990,	CAMBRIDGE MA. US	1,2,9, 11,12
		35 - 645;	ORIDITALLY 181, 00	,
	L. GEORG	GI ET AL.: 'daf-1, a C	C. elegans gene	
	control	ling Dauer larva devel	lopment, encodes a	
	novel re	eceptor protein kinase		
		n the application		
	see absi	tract; figures 4,6		
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^	COMMUNIC	CATIONS.		
	vol. 157	7, no. 2, 15 December	1988, DULUTH, MN,	
	US	\ \ \ \ \ \ \ \ \ \ \ \ \ \ \ \ \ \ \		
	pages 84	44 - 849; EN ET ALì: 'Characteri	tertion of activin A	
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NTS CONSIDERED TO BE RELEVANT (CONTINUED FROM THE SECOND SHEET)	
Citation of Document, with indication, where appropriate, of the relevant passages	Relevant to Claim No.
SCIENCE. vol. 255, no. 5052, 27 March 1992, WASHINGTON DC, US pages 1702 - 1705; L. MATHEWS ET AL.: 'Cloning of a second type of activin receptor and functional characterization in Xenopus embryos.' see abstract; figure 1	1-5, 9-12,20, 25
BIOCHEMICAL AND BIOPHYSICAL RESEARCH COMMUNICATIONS. vol. 184, no. 1, 15 April 1992, DULUTH, MN, US pages 310 - 316; C. DONALDSON ET AL.: 'Molecular cloning and binding properties of the human type II activin receptor.' see abstract; figure 2	1-5, 9-12, 16-20, 22-24
CELL vol. 68, no. 4, 21 February 1992, CAMBRIDGE, MA, US pages 775 - 785; H. LIN ET AL.: 'Expression cloning of the TGF-beta type II receptor, a functional transmembrane serine/threonine kinase.' see abstract; figures 2,5,6	1-3, 9-12, 22-24
BIOCHEMICAL AND BIOPHYSICAL RESEARCH COMMUNICATIONS. vol. 181, no. 2, 16 December 1991, DULUTH, MN, US pages 684 - 690; M. KONDO ET AL.: 'Activin receptor mRNA is expressed early in Xenopus embryogenesis and the level of the expression affects the body axis formation.' see abstract; figure 1	1-5, 9-12, 16-18, 20,22-24
	SCIENCE. vol. 255, no. 5052, 27 March 1992, WASHINGTON DC, US pages 1702 - 1705; L. MATHEWS ET AL.: 'Cloning of a second type of activin receptor and functional characterization in Xenopus embryos.' see abstract; figure 1 BIOCHEMICAL AND BIOPHYSICAL RESEARCH COMMUNICATIONS. vol. 184, no. 1, 15 April 1992, DULUTH, MN, US pages 310 - 316; C. DONALDSON ET AL.: 'Molecular cloning and binding properties of the human type II activin receptor.' see abstract; figure 2 CELL vol. 68, no. 4, 21 February 1992, CAMBRIDGE, MA, US pages 775 - 785; H. LIN ET AL.: 'Expression cloning of the TGF-beta type II receptor, a functional transmembrane serine/threonine kinase.' see abstract; figures 2,5,6 BIOCHEMICAL AND BIOPHYSICAL RESEARCH COMMUNICATIONS. vol. 181, no. 2, 16 December 1991, DULUTH, MN, US pages 684 - 690; M. KONDO ET AL.: 'Activin receptor mRNA is expressed early in Xenopus embryogenesis and the level of the expression affects the body axis formation.'

INTERNALIONAL SEARCH REPORT

International application No.

PCT/US 92/03825

Box	x I	Observations where certain claims were found unsearchable (Continuation of item 1 of first sheet)
Thi	s int	ernational search report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:
2.		Claims Nos.: because they relate to subject matter not required to be searched by this Authority, namely: Although claims 28 and 31 (both practically, as far as an in vivo method is concerned) are directed to a method of treatment of the human/animal body, the search has been carried out and based on the alleged effects of the compound/composition. Claims Nos.: because they relate to parts of the international application that do not comply with the prescribed requirements to such an extent that no meaningful international search can be carried out, specifically:
3. [Box		Claims Nos.: because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a). Observations where unity of invention is lacking (Continuation of item 2 of first sheet)
1.00	ine	rnational Searching Authority found multiple inventions in this international application, as follows: .
1. [As all required additional search fees were timely paid by the applicant, this international search report covers all searchable claims.
2. [As all searchable claims could be searches without effort justifying an additional fee, this Authority did not invite payment of any additional fee.
з. [As only some of the required additional search fees were timely paid by the applicant, this international search report covers only those claims for which fees were paid, specifically claims Nos.:
4. [)	No required additional search fees were timely paid by the applicant. Consequently, this international search report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:
Rem	ark o	The additional search fees were accompanied by the applicant's protest. No protest accompanied the payment of additional search fees.

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